



REVIEW

Open Access



Medicinal plants with hepatoprotective potentials against carbon tetrachloride-induced toxicity: a review

Chidiebere Emmanuel Ugwu * and Stephen Monday Suru

Abstract

Background: Carbon tetrachloride (CCl_4) is a well-characterized hepatotoxic agent. With rising cases of liver diseases, the identification, assessment, and development of hepatoprotective agents from plants source has become imperative.

Main body: With arrays of literature on plants with hepatoprotective potentials, this review sourced published literatures between 1998 and 2020 and systematically highlighted about 92 medicinal plants that have been reported to protect against CCl_4 -induced liver injury in animal models. The results show that herbal plants provide protection for the liver against CCl_4 by downregulation of the liver marker enzymes and activation of antioxidant capacity of the liver cells with the restoration of liver architecture. We also provided the traditional and accompanying pharmacological uses of the plants. A variety of phytochemicals mostly flavonoids and polyphenols compounds were suggested to offer protection against liver injuries.

Conclusion: It can be concluded that there are a variety of phytochemicals in plant products with hepatoprotective activity against CCl_4 -induced toxicity in animal models.

Keywords: Carbon tetrachloride, Medicinal plants, Hepatoprotective, Silymarin, Folkloric medicine

Background

The liver being an important organ is often exposed to array of threats [1]. Injury to the liver can lead to deterioration of its functions and may culminate in organ failure [2]. The likely risk factors for the development of the liver diseases have been suggested to include pathogenic microorganisms and viruses, hepatotoxins, overdose and duration of drugs, obesity and malnutrition, alcohol, autoimmune disorders, type-2 diabetes, and genetic factors [1]. The diseases of the liver are of public health concern because orthodox remedies for liver diseases produce limited results with attendant side effects. As such, utilization of complementary and alternative

herbal medicine has attracted research interest for novel plausible hepatoprotective agents capable of ameliorating or reversing liver injury with little side effects [3, 4]. Over the years, this search has gained impetus with many studies focusing on hepatoprotective potentials of plant drugs.

Carbon tetrachloride (CCl_4) is a known hepatotoxicant in humans and animal models [5]. It has been successfully used in hepatotoxicity research as a model and to appraise hepatoprotective agents [6, 7]. With reports on the rise of liver diseases and numerous literature reports on plants with potential hepatoprotective activity, this review highlighted the mechanism of CCl_4 toxicity, the significance, effectiveness, and underlying mechanisms of herbal plant extracts on CCl_4 -induced toxicity in experimental animal models.

*Correspondence: ce.ugwu@unizik.edu.ng

Department of Human Biochemistry, Faculty of Basic Medical Sciences, College of Health Sciences, Nnamdi Azikiwe University (Nnewi Campus), Nnewi, Anambra State, Nigeria

Main text

Insight on the mechanism of carbon tetrachloride hepatotoxicity

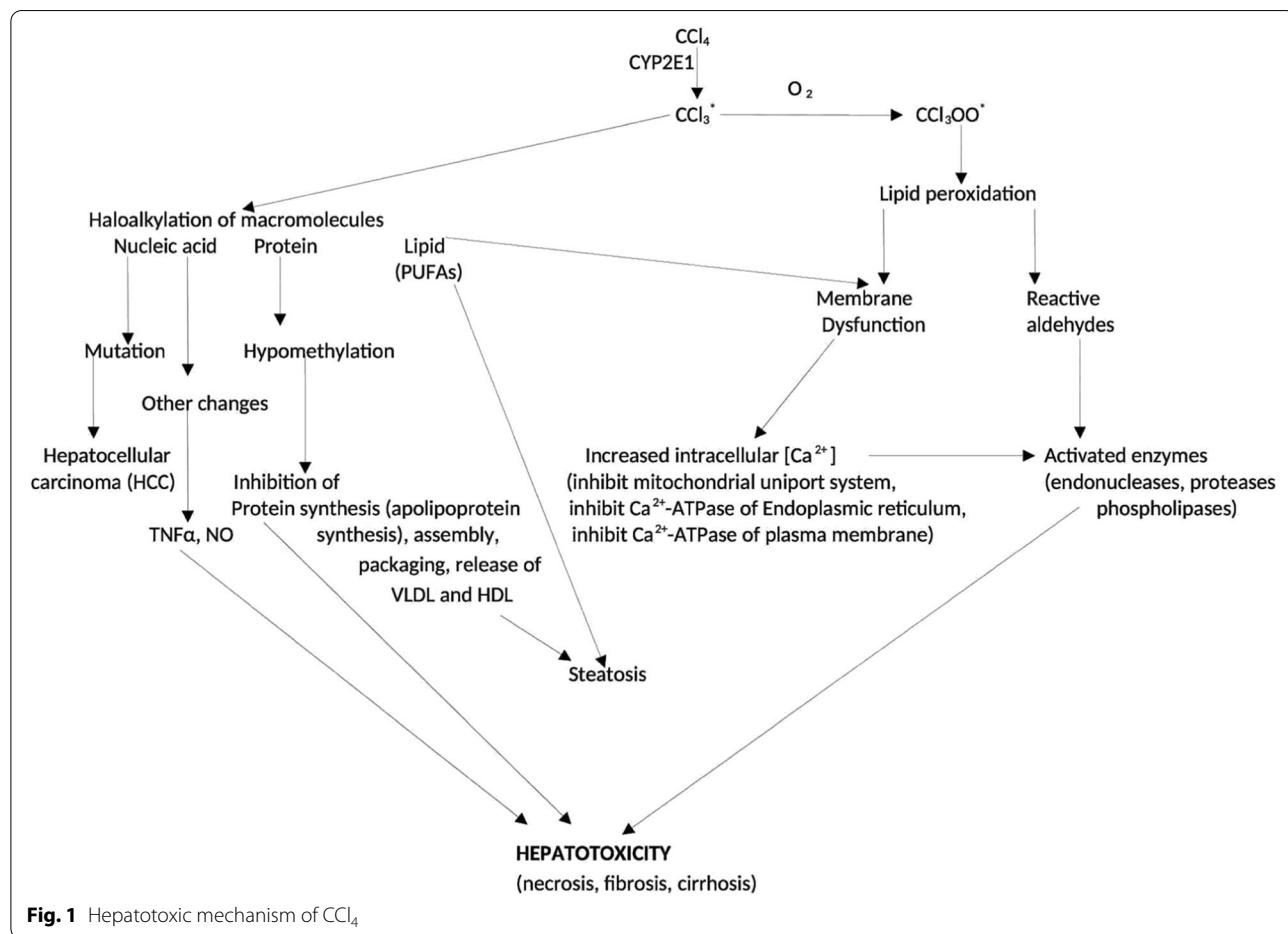
Prior to the Montreal Protocol, CCl_4 was formerly and widely used as a fire suppressant, as a precursor to refrigerants, propellants for aerosol cans, as a cleaning agent, a widely used solvent in organic chemistry, as a pesticide, and anesthetics [8, 9]. However, it is rarely used today because of adverse health effects and environmental safety concerns. Symptoms associated with acute inhalation of low–medium doses include headache, weakness, lethargy/general anesthesia, nausea, vomiting, and respiratory arrest. For medium to high oral exposure, the liver is known to be the primary site of CCl_4 -induced toxicity beginning with acute but progressive centrilobular injury that may culminate in cell death [10].

Experimental deductions

Due to the complex nature of CCl_4 -induced liver damage, there have emerged several independent mechanisms to explain each of the facets of the associated changes. The interrelationship among diverse mechanisms proposed

for each of these associated changes has not been well-established/outlined. This is primarily because early and later changes associated with the hepatotoxic development have been mixed up. As a result, a harmonized understanding of the intricate mechanisms involved in hepatic damage has become partly elusive. However, this has not obscured the following experimental deductions (Fig. 1):

- Changes in endoplasmic reticulum (ER) function due to decrease in glucose-6 phosphatase [11], which may not be unconnected with CCl_4 -induced glycogen depletion and attendant protection from carbohydrate-rich diets [12, 13]. Besides, CCl_4 -induced disruption and disassociation of polyribosomes from ER alters its anabolic function as manifested in decreased incorporation of amino acids into proteins such as albumin and fibrinogen [14]. Additionally, CCl_4 -induced hypomethylation of 2'-O-ribose moieties in rRNA might have resulted from transient increase in cytosolic Ca^{2+} . This increase may activate the selective destruction of rRNA methylases via



the action of demethylases or proteases. Overall, the protein synthetic function of ER in the centrilobular region may be hampered with an attendant defects in the ability of the liver to effectively respond to additional insults [10].

- Calcium homeostasis underlies some aspects of CCl_4 hepatotoxicity (plasma membrane blebbing and fatty accumulation- steatosis); CCl_4 may elicit dramatic redistribution of intracellular Ca^{2+} stores, albeit no total cellular change [10]. Calcium ion (Ca^{2+}) homeostasis is maintained by 3 mechanisms: (i) Ca^{2+} extrusion by plasma membrane ATPase, (ii) Ca^{2+} sequestration by mitochondria, and (iii) Ca^{2+} sequestration by liver ER. So, CCl_4 may cause decreased Ca^{2+} sequestration by ER and mitochondria, decreased extrusion by plasma membrane ATPase, as well as blockage of gap junctional intercellular communication may favor increase cytosolic Ca^{2+} . An ATP-dependent Ca^{2+} sequestration by hepatic ER has been shown to be disrupted by CCl_4 [15]. Endoplasmic reticulum membrane permeability may also be altered, being one indicator of impending cell death [16].
- Rapid destruction/decrease in cytochrome P₄₅₀ in centrilobular regions (suggesting that CCl_4 was metabolized by ER mixed-function oxidase system), which is orchestrated by low levels of reduced glutathione (GSH) and low oxygen tension. In turn, low level oxygen tension may limit competition between O₂ and CCl_4 for cytochrome P₄₅₀ binding (i.e., CCl_4 may readily bind to cytochrome P₄₅₀).
- Metabolic products [trichloromethyl (CCl_3^{\bullet}) or peroxytrichloromethyl ($\text{CCl}_3\text{-OO}^{\bullet}$) free radical] elicit damage: lipid peroxidation of vulnerable unsaturated fatty acids in membrane phospholipids and destruction of haem moiety of cytochrome P₄₅₀.
- Blockage of gap junctional communication by CCl_4 thereby shutting down intercellular communication.
- Changes in mitochondrial function: disruption of oxidative phosphorylation due partly to chelation of calcium [17].

Making sense out of experimental deductions

The hepatic biotransformation of CCl_4 primarily involves metabolic activation to transient reactive intermediates. Under low oxygen partial pressure, cytochrome P₄₅₀ catalyzes the reductive de-halogenation of CCl_4 resulting in predominant formation of CCl_3^{\bullet} and CHCl^{\bullet} radicals [18, 19]. These reactive intermediates may bind covalently to cellular components (membranes, microsomes) and impinge on mostly lipid metabolism (increased synthesis,

decreased transport out of the hepatocyte) thereby culminating in hepatic steatosis (fatty liver) [20, 21].

Dianzani [22] reported that covalent modification of lipoproteins occurs prior to their decreased transport out of hepatocytes. Intracellular maturation of lipoproteins in the Golgi apparatus is dependent on galactosylation which is catalyzed by glucosyl- and galactosyltransferases [23]. The CCl_4 -induced damage of Golgi apparatus and eventual reduction in the activities of these enzymes may explain the observed decrease in lipoprotein secretion associated with CCl_4 intoxication. Thus, CCl_4 -induced inhibition of lipoprotein secretion, and its attendant hepatic steatosis mainly result from covalent binding of CCl_4 metabolites to cell constituents, but not due to lipid peroxidation.

Under high oxygen partial pressure, however, CCl_3^{\bullet} may interact with oxygen to form $\text{CCl}_3\text{-OO}^{\bullet}$. The peroxy radicals may elicit the peroxidation of unsaturated fatty acids especially in membrane phospholipids of intracellular and plasma membranes [24]. Some of the lipid peroxidative products may inflict further damage leading to increased membrane permeability and a comprehensive loss in membrane integrity [25]. Thus, both covalent binding of CCl_4 metabolites and lipid peroxidation work in tandem to elicit the hallmark of damage seen in CCl_4 -induced hepatotoxicity.

The consequences of loss of membrane integrity are enormous and may lead to cascade of events culminating in liver necrosis. These events may include disturbed Ca^{2+} homeostasis/dramatic redistribution of Ca^{2+} in hepatocytes, leakage/efflux of K⁺, and influx of Na⁺ [10, 26].

Beside the peroxidative action, CCl_4 -derived free radicals and their attendant oxidative stress have been shown to enhance NF- κ B expression, which in turn initiates the synthesis of cytotoxic cytokines, which may be partly responsible for liver injury [27]. Tumor necrosis alpha (TNF- α) has been implicated in CCl_4 -induced hepatocellular damage [28]. At lower doses of CCl_4 , inflammatory responses prevail. Healthy hepatocytes are insensitive to tissue necrosis factor alpha (TNF- α) action, but become sensitive once protein and RNA synthesis are inhibited [29].

Summarily, CCl_4 hepatotoxicity may be due to a combination of factors such as the thorough inhibition of protein synthesis, the severe derailment of intracellular Ca^{2+} sequestration, and the effect on membrane integrity. These factors may result and progress through a series of steps that contribute to various extents to the ultimate damage: reductive dehalogenation, covalent binding of resulting radicals; inhibition of protein synthesis (in particular, apolipoprotein synthesis), assembly, packaging and release of VLDL and HDL, fat accumulation;

Table 1 List of traditional plants with anti-hepatotoxic potential against acute carbon tetrachloride hepatotoxicity

s/n.	Botanical name	Family	Plant part/extract	Folkloric use	Pharmacological use	Reference
1	<i>Abelmoschus manihot</i> (L.) medic	Malvaceae	Flower, ethanol	Treatment of jaundice and hepatitis, control of fertility, easing of child birth and stimulation of lactation.	Anti-inflammatory, antioxidant, antibacterial, anticonvulsant, cardioprotective, and neuroprotective actions	[32]
2	<i>Acacia mellifera</i>	Fabaceae	Leaves, acetate/aqueous/n-h-butanol	Treatment of cold, malaria, syphilis, and bowel problems.	Antimalarial, antimicrobial, antiviral activity against HIV-1, and herpes simplex virus	[33]
3	<i>Aegle marmelos correa ex Roxb</i>	Rutaceae	Pulp/seed, aqueous	Treatment of jaundice, hepatitis, piles, tuberculosis and antidiarrheal. Used as stomach tonic.	Antidiarrhoeal, anti-inflammatory, and wound healing effects	[34]
4	<i>Aegle marmelos correa ex Roxb with pipérine</i>	Rutaceae	Leaves, 70% ethanol	Used as astringent, laxative and expectorant. Treatment of inflammation, cataract, diabetes, diarrhea, and asthma.	Antifungal, ulcer healing, anti-inflammatory, antidiabetic, diuretic, anticancer, and antioxidant properties	[35]
5	<i>Alangium salvifolium</i> .	Alangiaceae	Stem bark, methanol.	Treatment of rheumatism, cancer and hemorrhoids. Root used to manage skin diseases, diarrhea, fever, carminative, and purgative expectorant.	Antiarthritis, androgenic, anthelmintic, antidiabetic, hepatoprotective, and anti-inflammatory effects	[36]
6	<i>Alhagi maurorum</i> (camel thorn)	Fabaceae	Leaves, methanol	As a remedy for rheumatic pains, biliarzias, liver disorders, and urinary tract infection.	Antioxidant antidiarrheal, and antilecerogenic activities.	[37]
7	<i>Alhagi maurorum</i> Medikus.	Fabaceae	Aerial parts, 90% ethanol	Treatment of liver problems, migraine and cataract. As a tonic digestive, antipyretic, laxative, diuretic, and aphrodisiac	Antiucler, antibacterial, antioxidant, anti-inflammatory, analgesic, antipyretic, antifungal, and hepatoprotective effects	[38]
8	<i>Allium sativum</i> (single clove garlic)	Amaryllidace	Garlic bulbs, 70% ethanol	Used as nutraceuticals	Antidiabetic, anticancer, antioxidant, immune modulation activities, and lowering of blood pressure.	[39]
9	<i>Amaranthus spinosus</i>	Amaranthaceae	Whole plant, 50% ethanol	Prevent swelling around the stomach. Used in the treatment of jaundice	Anti-inflammatory, antimarial, antibacterial, antidiuretic, antiviral, immunostimulatory, and antioxidant effects	[40]
10	<i>Amorphophallus campanulatus</i> (Roxb)	Araceae	Tubers, aqueous	Treatment of piles, abdominal pain, tumors, enlargement of spleen, asthma, and rheumatism	Antibacterial, antifungal, and cytotoxic activities	[41]
11	<i>Argemone Mexicana</i> L	Papaveraceae	Crude powder leaf	Treatment of malaria, fever, abdominal pains, and jaundice	Antibacterial, anti-inflammatory, wound-healing, antifertility, anti-stress, anti-allergic, cytotoxic, antidiabetic, and antihepatotoxic activities	[42]
12	<i>Artemisia iwayomogi</i>	Compositae	Aqueous	Treatment of hepatic disorders	Antioxidant, cytoprotection, choleretic hepatoprotection, antimicrobial, anti-inflammatory and antibiotic effects.	[43]

Table 1 (continued)

s/n.	Botanical name	Family	Plant part/extract	Folkloric use	Pharmacological use	Reference
13	<i>Bauhinia variegata</i>	Leguminosae.	Stem bark, alcohol	Treatment of bronchitis, leprosy, diarrhea, piles, and tumor. Used as astringent	Hypoglycaemic, haemagglutinating, antibacterial, and antifungal effects	[44]
14	<i>Bougainvillea spectabilis</i>	Nyctaginaceae	Esculetin	Treatment of liver damage, cough, pertussis, and bronchitis	Antimicrobial, anticancer, antiabetic, anti-inflammatory, antihyperlipidemic, antioxidant, antiulcer, and antihepatotoxic activities	[45]
15	<i>Bryonia dioica Jacq</i>	Cucurbitaceae	Leaves, 80% ethanol	Treatment of various inflammatory conditions, bronchial complaints, asthma, intestinal ulcer, hypertension, and arthritis. Applied as a rubefacient to muscular pains. Treatment of fever and bronchitis	Antinociceptive, antimicrobial, antioxidant, hepatoprotective, anticancer, hypercholesterolemia, analgesic, anti-inflammatory, cytotoxic, and hepatoprotective	[46]
16	<i>Bryoscarpus coCCheus Schum</i>	Connaraceae	Leaves, aqueous	Mouth and skin sores, swellings, tumors, earache, muscular pain, and jaundice	Antioxidant and hepatoprotection	[47]
17	<i>Cajanus cajan</i>	Leguminosae.	Aerial, 70% ethanol	Jaundice and stomach disorders	Anthelmintic, antioxidant and protection against alcohol-induced liver damage	[48]
18	<i>Calotropis gigantean R.Br</i>	Asclepiadaceae	Stem, 50% ethanol.	In tooth ache and ear ache, sprain, anxiety, pain, epilepsy, and in mental disorders	Antidiarrheal, analgesic, CNS activity, and pregnancy interceptive properties	[49]
19	<i>Camellia nitidissima Chi</i>	Theaceae	Leaves, 10 % ethanol	Treatment of dysentery, hypertension, diarrhea, faulcts, hepatitis, jaundice, liver cirrhosis and sores	Leaves show antioxidant, antimotor, antibacterial, anti-inflammatory, hypoglycaemic, hypolipidemic, antidepressant, antilergic, and immunomodulatory activities	[1]
20	<i>Canna indica L</i>	Cannaceae	Aerial part, methanol	Treatment of diuresis, fever, dropsey, earaches, and eye disease	Analgesic, antioxidant, and hepatoprotective effects	[50]
21	<i>Capparis spinosa</i>	Capparidaceae	Root bark, 80% ethanol	Treatment of hepatic diseases. Reducing flatulence, treatment of rheumatism, anemia, and gout. Used as diuretics	Antidiabetic, hypoglycaemic, antioxidant, apoptotic, antibacterial, anti-inflammatory, antifungal, and hepatoprotective effects	[51]
22	<i>Capsella bursa-pastoris (L.) Medik</i>	Brassicaceae	Aerial parts, 90% ethanol	Remedy for liver, hemorrhages, respiratory problem, and as diuretic	Antimicrobial, antioxidant, anticaner, anti-inflammatory, and sedative effects	[38]
23	<i>Carissa opaca</i>	Apocynaceae	Leaves, 95% methanol	Treatment of asthma, cardiac disorder and cough	Antioxidant membrane stabilization, antipyretic and aperient activities	[52]
24	<i>Carthamus tinctorius L</i>	Asteraceae	Flower, hydroxyafflor yellow A	Treatment of dysmenorrhea, amenorrhea, postpartum abdominal pains, and pains of the joints. As antidote to poisoning and purgative	Antioxidant antidiabetic, hepatoprotective, anti-inflammatory, antifungal, antimicrobial, and hepatoprotective effects	[53]

Table 1 (continued)

s/n.	Botanical name	Family	Plant part/extract	Folkloric use	Pharmacological use	Reference
25	<i>Carthamus tinctorius</i> . L	Asteraceae	Flower, Na ₂ CO ₃	Treatment of gynecological diseases, osteoporosis, cardiovascular diseases, and angitis	Nutraceutical, hepatoprotective, antioxidant, promoting blood circulation, and inhibiting platelet aggregation, anti-inflammatory, antipyretic, anti-tumor, and antidiabetic activities	[54]
26	<i>Carum carvi</i>	Apiaceae	Fruit, aqueous	Treatment of jaundice, indigestion and pneumonia. As appetizer, diuretic and gastric stimulant	Anti-inflammatory, spasmyolytic, antimicrobial, antioxidant, camimative, antidiabetic, immunomodulatory, anticancer, and hypolipidaemic properties	[55]
27	<i>Cassia angustifolia</i> Vahl	Caecalpiniaceae	Leaves, ethanol	Used in jaundice, rheumatoid arthritis, blood disease, diarrhea, ringworm, skin diseases, dysentery and as laxatives	Hepatoprotection and antioxidant activities	[56]
28	<i>Cassia angustifolia</i> Vahl	Leguminosae	Leaves, 90% alcohol	Used as laxative, febrifuge, treatment of anemia, typhoid, cholera, jaundice and tumors	Hepatoprotection and antioxidant activities	[57]
29	<i>Cassia fistula</i> Linn	Caesalpiniaceae	Leaves, 90% ethanol	Treatment of jaundice and rheumatism. Used as a laxative.	Hepatoprotective and antioxidant properties	[58]
30	<i>Cichorium intybus</i>	Asteraceae	Esculetin	Treatment of acne, inflammation of throat, jaundice, enlargement of spleen, diarrhea, vomiting, and rheumatism	Hepatoprotection, antimicrobial, antidiabetic, and analgesic effects	[45]
31	<i>Cichorium intybus</i>	Asteraceae	Seed, ethanol	Treatment of acne, inflammation of throat, jaundice, enlargement of spleen, diarrhea, vomiting, and rheumatism.	Hepatoprotection, antihelminthic, antimicrobial, antidiabetic, and analgesic effects	[59]
32	<i>Cichorium intybus</i>	Asteraceae	Seed, 0.03% methanol	Treatment of acne, inflammation of throat, jaundice, enlargement of spleen, diarrhea, vomiting, and rheumatism	Hepatoprotection, antihelminthic, antimicrobial antidiabetic, and analgesic effects.	[59]
33	<i>Cichorium intybus</i>	Asteraceae	Leaves, hydroethanol (1:1)	Treatment of acne, inflammation of throat, jaundice, enlargement of spleen, diarrhea, vomiting, and rheumatism	Hepatoprotection, antihelminthic, antimicrobial, antidiabetic and analgesic effects.	[60]
34	<i>Cinnamomum verum</i>	Lauraceae	Cinnamon powder, 95% ethanol	Treatment of diabetes, respiratory, and gynecological ailments	Enhancement of glycogen synthesis, antioxidant, antidiabetic, hypolipidemic, antipyretic, and analgesic activities	[61]

Table 1 (continued)

s/n.	Botanical name	Family	Plant part/extract	Folkloric use	Pharmacological use	Reference
35	<i>Cinnamomum verum</i>	<i>Lauraceae</i>	Bark essential oil, dichloromethane	Preventing heart diseases, reduction in cholesterol and as an antidiabetic	Antioxidant boosting cognitive activity, antiangiogenesis, anti-inflammatory, antimicrobial, and protection against Parkinson's disease	[62]
36	<i>Cinnamomum zeylanicum L</i>	<i>Lauraceae</i>	Bark, 80% ethanol	Flavoring for foods and in traditional medicine to treat variety of health conditions	Antimicrobial, insecticidal, antityrosinase, antioxidant, antimutagenic, anti-inflammatory, hypotensive, and cholesterol-lowering effects.	[63]
37	<i>Citrus aurantium</i> (essential oil)	<i>Rutaceae</i>	Peel skin, aqueous oil	Diaphoretic and antiseptic	Analgesic, anti-inflammatory, antifungal, and antibacterial activities	[64]
38	<i>Citrus limon</i> (L.) Burm.F	<i>Rutaceae</i>	Fruit, 70% ethanol	Treatment of liver ailment and jaundice. Treatment of sluggish liver, rheumatism, fever, and febrile diseases	Chenoprevention, lipid peroxidation inhibitor, hypocholesterolemic, and antioxidant effects.	[65]
39	<i>Clerodendrum volubile</i>	<i>Verbenaceae</i>	Leaves, 50% methanol.	Treatment of diabetes, ulcer, arthritis, and rheumatism	Antidiabetic, antihypertensive, antioxidant, and anticancer effects	[66]
40	<i>Clitoria ternatea L</i>	<i>Fabaceae</i>	Leaves, ethanol	Treatment of liver diseases, insect bites, asthma, leukoderma, and inflammation	Antihelmintic, antihistaminic, antimicrobial, cytotoxic, anti-inflammatory, wound healing, proteolytic, hypoglycemic, and antioxidant activities	[56]
41	<i>Coriandrum sativum. L</i>	<i>Apiaceae</i>	Leaves, ethanol	Treatment of jaundice	Anxiolytic, antidepressant and sedative, hypnotic effects. Neuroprotective, antibacterial, anti-inflammatory, analgesic, antidiabetic, antifungal, and hypolipidaemic effects	[67]
42	<i>Coriandrum sativum. L</i> (essential oil)	<i>Apiaceae</i>	Fruits, aqueous	Recommended for spastic condition of the gastro intestinal oral tract, flatulence, fullness and loss of appetite due to their antispasmodic, and antimicrobial activities	Anxiolytic, antidepressant and sedative-hypnotic effects. Neuroprotective, antibacterial, anti-inflammatory, analgesic, antidiabetic, antifungal, and hypolipidaemic effects	[55]
43	<i>Coriandrum sativum</i>	<i>Umbellifera</i>	Leaves/stem, 70% ethanol	Treatment of ailments like spasm, rheumatism, neuralgia, gastric complaint, bronchitis, diarrhea, carminative and diuretic tonic	Hypoglycemic, antibacterial, antifungal, free radical scavenging, and lipid peroxidation properties	[68]
44	<i>Cortex dictamni</i>	<i>Rutaceae</i>	Whole plant, aqueous	Treatment of Jaundice, chronic hepatitis, cough rheumatism and some skin diseases. To clear heat, dry dampness, dispel wind, treatment of arthritis, eczema, rubella, and urticaria	Good scavenger of free radicals and inhibition of lipid peroxide	[69]

Table 1 (continued)

s/n.	Botanical name	Family	Plant part/extract	Folkloric use	Pharmacological use	Reference
45	<i>Curcuma longa</i> L	Zingiberaceae	Rhizome(oot), 50% ethanol and curcumin	Used for the treatment of chronic diseases like diabetes mellitus, dermatological infection, and depression	Anti-inflammatory, immunoregulatory, and antioxidant effects [70]	
46	<i>Cytisus scoparius</i> L	Leguminosae	Aerial, 70% ethanol	As a diuretic hypnotic, sedative, and antidiabetic	Used as diuretic, hypnotic, sedative, antidiabetic, and hepatoprotector [71]	
47	<i>Dicoma anomala</i> Sond	Asteraceae	Root, aqueous	Treatment of cold and cough, fever, ulcer, and dermatosis	Antispasmodial, antibacterial, anthelmintic, antiviral, antioxidant, and anti-inflammatory effects [72]	
48	<i>Dioscorea alata</i> peel	Dioscoreaceae	Peel, aqueous	To strengthen stomach function, anorexia, and to eliminate diarrhea	Anti-inflammatory effect [73]	
49	<i>Eclipta alba</i> (L) Hassk	Asteracea	Leaves, aqueous	Treatment of Jaundice. Juice used in treatment of hair problem, typhoid, dysentery and skin diseases	Hepatoprotection, antidiabetic, analgesic, antimicrobial, antioxidant, anticancer, anti-inflammatory, and immunoregulatory activities [74]	
50	<i>Emilia officinalis</i> (Gaertn)	Euphorbiaceae	Fruit, methanol	Relieving cough and skin diseases	Antidiabetic, cytoprotective, anti-ulcerogenic, immunomodulatory, antioxidant, and anticataractogenic effects [75]	
51	<i>Entada purpurea</i> DC	Fabaceae	Stem, 85% ethanol	Used as narcotic. Treatment of jaundice. As an anthelmintic, in curing eye diseases, diarrhea, and skin diseases	Hepatoprotective and antioxidant effects [76]	
52	<i>Ephedra foliate</i> Boiss	Ephedraceae	Aerial parts, 90% ethanol	Treatment of allergies, asthma, lung congestion, chills and cold	Antidiabetic, anticancer, antimicrobial, antioxidant, anti-inflammatory, and hepatoprotective effects [38]	
53	<i>Euphorbia dracunculoides</i> L	Euphorbiaceae	Aerial part, 95% methanol	Curing skin disorders and edema. Used as diuretic and laxative and in the treatment of rheumatism, snake bite and edema	Anti-inflammatory, analgesic and antioxidant activities. Hepatoprotection against hepatocyte cell lines [5]	
54	<i>Fagonia schweinfurthii(Hadid)</i> Hadid	Zygophyllaceae	Whole plant, ethanol.	Treatment of Jaundice, diabetes, joint pains, asthma and dropsy.	Antioxidant hepatoprotective, anti-inflammatory, wound healing and analgesic activities. Cytotoxic, hypoglycemic and antihelmintic activities. [77]	
55	<i>Ficus carica</i> Linn	Moraceae	Leaves, ethyl acetate.	Treatment of vitiligo, diabetes, cough, asthma, constipation and gingivitis.	Cytotoxic, hypoglycemic and antihelmintic activities. Treatment of rheumatism, arthropathy, chronic nephritis, menalgia, and menopausal syndrome. [78]	
56	<i>Flemingia macrophylla</i>	Fabaceae/Leguminosae	Root, aqueous.		Antioxidative, anti-inflammatory, analgesic, hypnotic-sedative and anxiolytic effects. [79]	

Table 1 (continued)

s/n.	Botanical name	Family	Plant part/extract	Folkloric use	Pharmacological use	Reference
57	<i>Ginkgo biloba</i>	Ginkgoaceae	Leaves, aqueous.	Treatment of Alzheimer's dementia and other cognitive dysfunctions.	Antioxidant cardioprotective, antiasthmatic, antidiabetic, management of cerebral insufficiency, and decreased gastric injury caused by ethanol.	[80]
58	<i>Glyphaea brevis</i>	Tiliaceae	Leaves, 50% methanol.	Treatment of hepatitis, jaundice and impotence.	Carminative, anticonvulsant effects, anti-inflammatory, antioxidant and improvement of lipid metabolism.	[81]
59	<i>Graptophyllum paraguayense E. Walther</i>	Crassulaceae	Leaves, aqueous	Regulation, alleviation of hepatic disorders, relief of pain, detumescence and carbuncles	Antioxidant anti-inflammatory, neuroprotective, hypotension regulation, antioxidant activity, and inhibition of cancer cells	[82]
60	<i>Hibiscus sabdariffa L.</i>	Malvaceae	Aerial parts, 90% ethanol	Used to prepare herbal drinks and as a flavoring agent. As diuretic and choleric	Antibacterial, antioxidant, nephro-protective, antidiabetic and antihypertensive effects	[38]
61	<i>Hippophae rhamnoides L</i>	Elaeagnaceae	Seabuckthorn berry polysaccharide, alcohol.	Treatment of asthma and circulatory disorders	Antioxidative, antimicrobial, antithrombotic, cardioprotective, hepatoprotective, radioprotective, and anti-inflammatory effects	[83]
62	<i>Indigofera oblongifolia</i>	Leguminaceae	Whole plant, 90% ethanol	Treatment of hepatic diseases and dysentery; enlargements of liver and spleen. An antidote of poison	Antimicrobial, anti-inflammatory and analgesic activities	[84]
63	<i>Launaea procumbens</i>	Asteraceae	Aerial parts, chloroform	Treatment of kidney disorders, hormonal imbalance, and sexual diseases	Spasmogenic, cardiovascular, anti-carcinogenic, anti-inflammatory, hepatoprotective, and antioxidant properties	[85]
64	<i>Lawsonia intermis L (Henna)</i>	Lythraceae	Leaves, 99% methanol	Used as astringent, hypotensive, sedative against headache. Treatment of jaundice, leprosy, and nervous disorder	Antimicrobial, anti-tumorigenic, anti-inflammatory, anti-apoptotic, antihyperglycaemic, antidiabetic, antiviral, and hepatoprotective effects	[86]
65	<i>Lawsonia intermis Linn</i>	Lythraceae	Leaves, aqueous	Treatment of liver diseases, jaundice, and burn	Anti-inflammatory, antipyretic, analgesics, antimicrobial, anticancer, and hepatoprotective properties	[87]
66	<i>Leucas cephalotes Linn.</i>	Labiatae	Whole plant, methanol	Treatment of liver disease, snake bite, and bronchitis, inflammation and jaundice.	Antifilarial and antidiabetic activities.	[88]
67	<i>Lobularia maritima</i>	Brassicaceae	Leaves, 10% ethanol	Antiscorbutic, diuretic, and as an astringent	Antioxidant and anti-inflammatory effects	[7]

Table 1 (continued)

s/n.	Botanical name	Family	Plant part/extract	Folkloric use	Pharmacological use	Reference
68	<i>Luffa acutangula</i> (Var.) amara	Cucurbitaceae	Leaves, ethanol	As a laxative and carminative digestible. Treatment of anemia, jaundice, biliousness, bronchitis, asthma, and piles	CNS depressant, antioxidant, and larvical activities	[89]
69	<i>Lygodium flexuosum</i> (L.) Sw	Lygodiaceae	Whole plant, n-hexane	Treatment of jaundice and liver disorders	Hepatoprotection against CCl ₄	[90]
70	<i>Madhuca indica</i> Syn	Sapotaceae	Bark, methanol	Used as stimulants, demulcent, astringents, remedy of itching, and swelling	Anti-inflammatory analgesic, hepatoprotective, antipyretic, anti-hyperglycemic, antiulcer, and antidiabetic effects	[91]
71	<i>Madhuca indica</i> Syn	Sapotaceae	Leaves, 70% ethanol, 90% ethanol	Treatment of piles, emetic, laxative tonic, anti-burn, and wound healing	Antidiabetic, anti-inflammatory, analgesic, anti-pyretic, antishmatic, antiulcer, anticancer, hepatoprotective, and antibacterial effects	[92]
72	<i>Mahonia oiwakken Hayata</i>	Berberidaceae	Root, 90% ethanol	Rheumatthritis, dysentery, hepatitis, antidote, and antiphlogistic agent	Hepatoprotection, antioxidant, and anti-inflammatory	[3]
73	<i>Mallotus philippensis</i> Muell-Arg	Euphorbiaceae	Leaves, methanol	Treatment of jaundice, threadworm, hookworm, and roundworm infections. As a purgative and carminative	Anticestodal, antibacterial, wound healing, antifilarial, antioxidant, anti-inflammatory, and immunoregulatory effects	[93]
74	<i>Memondica tuberosa</i> Cogn	Cucurbitaceae	Tubers, 70% ethanol	Used as abortifacient	Antioxidant, antihyperglycemic, anticonvulsant, anti-inflammatory, antiovulation, antidiarrhoeal, and nephroprotective activities	[94]
75	<i>Mentha piperita</i> L	Lamiaceae	Leaves (essential oil)	Treatment of nausea, bronchitis, flatulence, liver complaints, ulcerative colitis, and as carminative	Antioxidant and anti-inflammatory	[95]
76	<i>Mentha arvensis</i> Linn	Lamiaceae	Leaves, aqueous, chloroform, ethanol	Carminative, antispasmodic, and anti-peptic ulcer agent	Radio protective, antispasmodic, antibacterial, anthelmintic, antifertility, hepatoprotective, antiulcer, and anti-inflammatory	[96]
77	<i>Mimosa pudica</i> 2009	Fabaceae/leguminosae	Leaves, methanol	Treatment of piles, fistula, insomnia, traumatic injury and jaundice	Hyperglycemic, antioxidant, anti-hepatotoxic, antidiabetic, wound healing, anti-inflammatory, and antimicrobial effects	[97]
78	<i>Mimosa pudica</i> Linn	Fabaceae/leguminosae	Leaves, ethanol	Treatment of wound, oedema, allergy, fever, diabetes, and indigestion	Hyperglycemic, antioxidant, anti-hepatotoxic, antidiabetic, wound healing, anti-inflammatory, and antimicrobial effects	[98]
79	<i>Momordica dioica</i> Roxb	Cucurbitaceae	Leaves, ethanol	Treatment of jaundice, hepatic diseases, fever, asthma, and as antihelminthic. Used as stomach laxative	Hypoglycemic, gastroprotective, ulcer healing, and hepatoprotective effects	[99]

Table 1 (continued)

s/n.	Botanical name	Family	Plant part/extract	Folkloric use	Pharmacological use	Reference
80	<i>Nerium oleander</i> Linn	Apocynaceae	Flower, methanol	Treatment of malaria and venereal diseases. Used as diuretic, insecticide, abortifacient, and cardiotonic. Relieves Indigestion	Cardiac insufficiency, anticonvulsant, antitumor, and antioxidant effects [100]	
81	<i>Nicotiana plumbiginifolia</i> L.	Solanaceae	Whole plant, methanol	Treatment of cuts, wounds, toothache, and rheumatic swelling	Antispasmodic, leaves are effective laxative, antioxidant, and antimicrobial [101]	
82	<i>Nymphaea alba</i> L.	Nymphaeaceae	Leaves, 76% ethanol	Used as antiseptic, an astringent and as a rubefacient in insomnia	Antioxidant, anti-inflammatory, and hepatoprotective effects. [6]	
83	<i>Olea europaea</i> L.	Oleaceae	Leaves, 20% oleuropein	Treatment of malaria and associated fever	Antimicrobial, anti-inflammatory, antioxidant, blood pressure lowering, lipid lowering, anticancer, and cardioprotective activities [102]	
84	<i>Origanum vulgare</i> .	Lamiaceae	Leaves, aqueous	Treatment of respiratory disorders, indigestion, and rheumatoid arthritis	Antihyperglycaemic, anti-inflammatory, cytotoxic, antioxidant, antithrombin, antimutagenic, and anti-carcinogenic effects [103]	
85	<i>Persea Americana</i> mill	Lauroceras	Leaves, aqueous	Remedy for pyorrhea. Toxic to silkworms	Antifungal, hypotensive, anti-inflammatory, anticonvulsant, antidiabetic, antioxidant, and vasorelaxant effects [104]	
86	<i>Phyllanthus niruri</i>	Phyllanthaceae	Aerial part, 80% ethanol	Treatment of urinary and bladder disorders, hepatic disorders, dyspepsia, influenza jaundice, and kidney stone	Hepatoprotective, antioxidant, antihyperuricemic, and lipid lowering effects [105]	
87	<i>Physalis peruviana</i> (Golden berry)	Solanaceae	Leaves, 50% methanol	Used as antispasmodic, diuretic, antiseptic, sedative, analgesic, and hepatitis	Antidiabetic, antimicrobial, anti-inflammatory and antihypercholesterolemic activities [106]	
88	<i>Pterogynium timorense</i> (DC) Leenh	Anacardiaceae	Bark, 70% methanol	–	Antimicrobial, hepatoprotective, antioxidant, anti-inflammatory, hypoglycemic, and cytotoxic effects [107]	
89	<i>Pleurotus ostreatus</i>	Pleurotaceae	Whole mushroom, 95% ethanol	Preventing heart disease, reduction in cholesterol, and treatment of diabetes	Inhibition of platelet aggregation, reduction of blood glucose and cholesterol, antibacterial, viral, and parasitic pathogens, and antioxidant activities [108]	
90	<i>Polygonum cuspidatum</i> sieb et Zucc	Polygonaceae	Rhizome, methanol	Treatment of jaundice, and to clear heat toxin, to promote blood circulation. Dispel stasis, suppress cough, and treat snake bites	Antidiabetic, anti-hepatitis B virus, antibacterial, anti-inflammatory, and antioxidant properties [4]	
91	<i>Premna esculetana</i> Roxb	Verbenaceae	Leaves, 95% ethanol	Treatment of hepatocellular jaundice, gout, hook worm infection, and snake bite	Antihyperlipidemic, hepatoprotective, antioxidant, analgesic, and anti-inflammatory activities [109]	

Table 1 (continued)

s/n.	Botanical name	Family	Plant part/extract	Folkloric use	Pharmacological use	Reference
92	<i>Raphanus sativus</i>	<i>Brassicaceae</i>	Leaves, aqueous and ethanol	Treatment of indigestion, abdominal bloating, diarrhea, bronchitis, intestinal parasites, and asthma	Antimicrobial, anticancer, antidiabetic, gastrointestinal, uterine tone modulatory, and cardio-modulatory activities	[110]
93	<i>Rourea induta planch</i>	<i>Connaraceae</i>	Leaves, 99% ethanol	Treatment of respiratory and kidney diseases. Treatment of blood diarrhea, and as diuretics	Anti-inflammatory hepatoprotective, antioxidant, and antipyretic activities	[111]
94	<i>Rubia cordifolia Linn</i>	<i>Rubiaceae</i>	Root, 50% ethanol	Treatment of jaundice	Potent antioxidant property, inhibit lipid peroxidation, anti-inflammatory, immunomodulatory, anticonvulsant, anxiolytic and antitumor activities	[112]
95	<i>Rumex vesicarius L</i>	<i>Polygonaceae</i>	Whole plant, methanol	Aperients, diuretic and cooling agent. Treatment of jaundice and dysentery. Curing stomach heat, toothache, and to promote appetite	Antimicrobial, anti-inflammatory, antioxidant, wound healing, and antitumor activities	[113]
96	<i>Semen celosia Cristatae L</i>	<i>Amaranthaceae</i>	Dry seeds, 60% ethanol	Treatment of hypertension, palsy, cataract, keratitis, diabetes, iridocyclitis, caligo corneal, and sarcoptidosis	Antibacterial, anticancer, antidiarrheal and anti-inflammatory effects	[114]
97	<i>Solanum trilobatum Linn</i>	<i>Solanaceae</i>	Whole plant, 90% ethanol	Used as an expectorant in the treatment of respiratory diseases, asthma, tuberculosis, and liver diseases	Broad spectrum antibiotic, antibacterial, antimitotic, anticancer, and antioxidant properties	[115]
98	<i>Solanum xantholoprum</i>	<i>Solanaceae</i>	Fruit, 50% ethanol	Laxative, treatment of enlargement of liver, antihelmintic, antipyretic, anti-inflammatory, antiasthmatic, and aphrodisiac activities.	Antiasthmatic, anti-nociceptive, antifungal, molluscicide, antispasmodic, antitumor, cardiotonic, hypotensive, antianaphylactic, and anti-ulrolithatic activities	[116]
99	<i>Spondias mombin</i>	<i>Anacardiaceae</i>	Leaves and stem, 50% methanol	Treatment of hepatitis	Antimicrobial, antiviral, anti-inflammatory, antihelminthic, hematinic sedative, antioxidant, and hepatoprotective effects	[117]
100	<i>Stachys pilifera Benth</i>	<i>Lamiaceae</i>	Leaves, 70% ethanol	Treatment of asthma, rheumatoid arthritis, and asthma	Anti-inflammatory, antioxidant, antibacterial, antitumor, and antimicrobial effects	[118]
101	<i>Vitis thunbergii Var</i>	<i>Vitaceae</i>	Aerial part, ethanol	Treatment of hepatitis, jaundice, diarrhea, and arthritis	Antioxidant, anti-inflammatory, antihypertensive, neuroprotective, antibacterial, and inhibition of adipocyte differentiation	[119]
102	<i>Xylaria nigripes(Koltz) Sacc</i>	<i>Xylariaceae</i>	Solid cultured mycelia, aqueous	Treatment of insomnia, trauma, diuretic, and nerve tonic	Antioxidant and hepatoprotective effects	[120]

Table 1 (continued)

s/n.	Botanical name	Family	Plant part/extract	Folkloric use	Pharmacological use	Reference
103	<i>Zingiber officinale</i> (Roscoe) rhizome (ginger)	Zingiberaceae	Rhizome, 90% methanol	Nutraceutical. Treatment of stomach aches, nausea, diarrhea, as carminative, appetite stimulant, and choleric	Antioxidant, anti-inflammatory, antitumor, antidiabetic, antimicrobial, neuro-protective, and gastro-protective potentials	[121]
104	<i>Zizyphus jujube</i> Mill	Rhamnaceae	Fruit, 70% ethanol	Invigorating the spleen, treatment of anorexia, lassitude, and control of hepatitis	Antioxidant and anti-inflammatory activities	[122]

Table 2 In vivo studies on medicinal plants with hepato protection against acute tetrachloride toxicity

s.no.	Botanical name	Animal model.	Maximum extract dose/route of administration	CCL4 dose/route of administration	Standard drug administered/route of administration	Result.	Active components.	Reference
1	<i>Abelmoschus manihot (L) medic</i>	Ku-Ming mice.	500mg/kg/b.w. (oral). 0.1 ml/kg/bw(0.12% v/v olive oil), i.p.	Biphenyl dicarboxylate (BDP) 150 mg/kg/b.w., oral.	ALT, AST, ALP, γ-GT, TNF-α, IL-1β, NO, MDA↓, GSH, SOD, GPx, CAT, GST↑.	Flavonoids, quercetin, hyperin, isquercetin, quercetin-3'-O-glucoside, nobiletin, myricetin	[32]	
2	<i>Acacia mellifera</i>	Wistar rats	500mg/kg/b.w.	1.25 ml/kg/b.w. (1:1 liquid paraffin) i.p	Silymarin, 100 mg/kg/bw	ALT, AST, GGT, ALP, TBL↓, P↑MDA, NP-SH ↑, Nonprotein TG↓, NP-SH↑. (Nonprotein sulfhydryl)	Flavonoids, saponin, tannins, triterpenoids	[33]
3	<i>Aegle marmelos correa ex Roxb</i>	Albino Wistar rats	- (oral)	0.2 ml/100g/b.w. (olive oil), i.p.	Silymarin 200 mg/kg/b.w. (oral)	AST, ALT, ALP, TB↓	Flavonoids	[34]
4	<i>Aegle marmelos correa ex Roxb</i>	Wistar albino rats	50mg/kg/b.w. (oral)	3ml/kg/b.w., i.p.	Silymarin 200 mg/kg/b.w. (oral)	ALP, ALT, AST, TB, LDH, MDA↓, SOD, CAT, GR, GSH, GST, GPx, G6PD, T.P↑, IL-10, TNF-α↓	Rutin, piperine	[35]
5	<i>Alangium salviifolium</i>	Swiss albino mice	50mg/kg/b.w. (oral)	1 ml/kg/b.w. (1:1 in olive oil).	—	AST, ALT, ALP, MDA, LDH, CYT-P450 reductase, cyt b5 reductase↓, SOD, CAT, DT-diaphorase, glutathione-S-transferase↑, ALT, AST↓	Piperine, γ-sitosterol	[36]
6	<i>Althagi maurorum (camel thorn)</i>	Wistar rats	660 mg/kg/b.w. (oral)	1 ml/kg/b.w. (maize oil) oral.	—	SGOT, SGPT, ALP, TB.	Flavonoids, phenols	[37]
7	<i>Althagi maurorum Medicus</i>	Wistar rats	500 mg/kg/b.w.	0.125 ml/kg (liquid paraffin, 1:1) i.p	Silymarin, 10 mg/kg (oral)	ALT, AST, ALP, TBL, TP↑, MDA↓, CAT, GST, SOD↑	Flavonoids, tannins.	[38]
8	<i>Allium sativum (Single clove garlic)</i>	Male rabbits	0.8 g (oral)	3 ml/kg/b.w. (1:1, olive oil)	—	AST, ALT, ALP, TB, MDA↓, GSH, SOD, CAT↑	—	[39]
9	<i>Amaranthus spinosus</i>	Sprague-Dawley rats	400 mg/kg/b.w. (oral)	1 ml/kg/b.w. (v/v olive oil) i.p.	Silymarin, 50 mg/kg/b.w. (oral)	MDA, Hydroperoxides↓, GSH, SOD, CAT↑	Flavonoids, phenols, betalains.	[40]
10	<i>Amorphophallus com-pulnatus (Roxb)</i>	Wistar albino rats and mice	500 mg/kg/b.w. (oral)	1 ml/kg/b.w. oral.	Silymarin, 100mg/kg (oral)	SGOT, SGPT, ALP Total bilirubin↓	Flavonoids	[41]
11	<i>Argemone Mexicana L</i>	Wistar rats	500 mg/kg/b.w. (oral)	0.5 ml/kg/b.w. i.p.	—	ALT, AST, ALP, MDA↓, TAC, GSH, SOD↑, Hydroxy proline↓	Leutolin, quercetin, queretin	[42]
12	<i>Artemisia iwayomogi</i>	Sprague-Dawley rats.	500 mg/kg/b.w (oral oliveoil) i.p.	2 ml/kg/b.w. (50% oliveoil) i.p.	—	AST, ALT, ALP, GGT↓, T.P↑, Total lipid↓	Scoparone	[43]
13	<i>Bauhinia variegata</i>	Sprague-Dawley rats	200 mg/kg/b.w. (oral)	1 ml/kg/b.w. (liquid paraffin, 1:1) subcutaneous.	—	—	—	[44]
14	<i>Bougainvillea spectabilis</i>	Wistar rats.	6 mg/kg/b.w. (oral)	—	AST, ALP, ALT↓	Esculetin	[45]	
15	<i>Byonia dioica Jacq</i>	Wistar albino rats	250 mg/kg/b.w. (gavage)	1.5 ml/kg, oral. 1 ml/kg/b.w. (corn oil, 1:1/v/v).	AST, AST↓	Flavonoids, terpenoids	[46]	

Table 2 (continued)

s.no.	Botanical name	Animal model	Maximum extract dose/route of administration	CCL4 dose/route of administration	Standard drug administered/route of administration	Result.	Active components.	Reference
16	<i>Bryoscarpus coCineus</i>	Albino rats	1000 mg/kg/b.w. (oral).	0.7 ml/kg/b.w. (1:1 in olive oil) i.p.	Ivoxin®, 200 mg/kg/b.w. (oral)	ALT, AST, ALP, MDA↓, TP, Albumin, CAT, SOD, GPx, GSH↑	Alkaloids, flavonoids	[47]
17	<i>Cajanus cajan</i>	Wistar albino rats	400 mg/kg/b.w. (oral)	2 ml/kg/b.w. (1:1 liquid paraffin), oral.	Liv 52, 100 mg/kg/b.w. (oral)	AST, ALT↓, T.P.↑	Alkaloid, flavonoids	[48]
18	<i>Calotropis gigantean R.Br.</i>	Wistar rats	500 mg/kg/b.w. (oral)	2 ml/kg/b.w. (1:1 olive oil), subcutaneous.	Silymarin, 100 mg/kg/b.w. (oral)	AST, ALT, LPO↓, GSH, SOD, GPx, CAT↑	Calotropin D1 and DII, calotropin F1 and FII.	[49]
19	<i>Camellia nitidissima Chi.</i>	Sprague-Dawley rats	160 mg/kg/day (i.p)	2 ml/kg (50% v/v, olive oil), i.p.	Thiopronin 20 mg/kg/day, (i.p)	AST, ALT, MDA↓, GSH, SOD↑, TNF-α, IL-6, IL-1β, NF-κB signaling↓, Nrf2 signaling pathway, HO-1, SOD, GSH↑	Polyphenols, flavonoids	[1]
20	<i>Canna indica L.</i>	Sprague-Dawley rats	200 mg/kg/b.w. (oral)	1.0 ml/kg/liquid paraffin, 1:2) i.p.	Silymarin, 25 mg/kg, (i.p)	SGPT, SGOT, ALP, TB, LP↓, GSH, CAT, T.P.↑	Lutein	[50]
21	<i>Capparis spinosa</i>	Mice	400 mg/kg/b.w. (oral)	0.2 ml/kg (olive oil 1:1), oral.	—	ALT, AST↓	Flavonoids, phenols, rutin, quercetin-3-O-glucoside, kaempferol, 3-O-rutinoside	[51]
22	<i>Capsella bursa-pastoris (L.) Medik.</i>	Wistar rats	500 mg/kg/b.w.	0.125 ml/kg (liquid paraffin, 1:1), i.p	Silymarin, 10 mg/kg, (oral)	SGOT, SGPT, ALP, TB	—	[38]
23	<i>Carissa opaca</i>	Sprague-Dawley rats	200 mg/kg/b.w. (intragastrically)	0.5 ml/kg/b.w. (20% v/v olive oil), i.p.	Silymarin, 50 mg/kg/b.w., (intragastrically)	AST, ALT, ALP, LDH, γ-GT↓, GSH-Px, GSR, SOD, GST, CAT, Peroxidase, Quinone reductase(QR)↑, TBARS, GSH, H2O2↓, T.P.↑	Isoquercetin, hyperoside, vitexin, myricetin, kaempferol	[52]
24	<i>Carthamus tinctorius L</i>	Sprague-Dawley rats.	5 mg/kg/day	1.0 ml/kg (olive oil).	—	ALT, AST, Hydroxy proline↓	Hydroxysafflor yellow A, isocarthamidin, carthamin, luteolin	[53]
25	<i>Carthamus tinctorius L</i>	Sprague-Dawley rats	20 mg/kg/b.w. (oral)	2 ml/kg/b.w. (1:1 olive oil), i.p.	Silymarin, 50 mg/kg/b.w. (oral)	ALT, AST, ALP, TP↓, Nrf2, GST, NQO1 expression, GSH↑, TBARS↓, SOD, CAT↑.	Carthamin, carthamidin, polyphenols, carthamus red, flavonoids	[54]
26	<i>Carum carvi</i>	NMR mice	0.13 g/kg/b.w. (oral)	2 ml/kg/b.w. (olive oil, 1:2), i.p.	—	AST, ALT, LP↓, GSH, GSH-Px↑, Px, XOD↓, Protein↑	Carvon	[55]
27	<i>Cassia angustifolia Vahl</i>	Wistar albino rats	300 mg/kg/b.w (oral)	2.5 ml/kg/b.w.	Silymarin, 100 mg/kg/b.w. (oral)	AST, ALT, ALP, Acid phosphatase(ACP), LDH, T.B., T.P.↑	Flavonoid, terpenoids, tannin, steroid	[56]
28	<i>Cassia angustifolia vahl</i>	Wistar rats	500 mg/kg/b.w. (oral)	4 ml/kg/b.w. (50% olive oil) oral	—	T.B, GOT, GPT↓, TP, GSH↑, LPO↓	Flavonoids	[57]

Table 2 (continued)

s.no.	Botanical name	Animal model	Maximum extract dose/route of administration	CCL4 dose/route of administration	Standard drug administered/route of administration	Result.	Active components.	Reference
29	<i>Cassia fistula Linn</i>	Wistar albino rats	500 mg/kg/b.w. (oral) paraffin)	0.1 ml/kg/b.w. (liquid)	-	MDA, AST, ALT, GSH, ALP, LDH, γ-glutamyltranspeptidase↓	Flavonoids	[58]
30	<i>Cichorium intybus</i>	Wistar rats	6 mg/kg/b.w. (oral)	-	AST, ALP, ALT↓	Esculetin	Cichotyboside	[45]
31	<i>Cichorium intybus</i>	Albino wistar rats	500 mg/kg/b.w. (oral) 50%, i.p.	1.5 ml/kg (olive oil 50%), i.p.	SGOT, SGPT, ALP↑, TP, albumin↑	SGOT, SGPT, ALP↑, TP, albumin↑	Cichotyboside	[59]
32	<i>Cichorium intybus</i>	Albino wistar rats	500 mg/kg/b.w. (oral)	1.5 ml/kg olive oil(50%), i.p.	SGOT, SGPT, ALP↑, TP, albumin↑	SGOT, SGPT, ALP↑, TP, albumin↑	Cichotyboside	[59]
33	<i>Cichorium intybus</i>	Albino rats	500mg/kg/b.w. (oral)	1.0 ml/kg olive oil 50%, i.p.	AST, ALP, ALT, TB↓, TP, albumin↑	AST, ALP, ALT, TB↓, TP, albumin↑	Esculetin and cichoty- boside	[60]
34	<i>Cinnamomum verum</i>	Wistar albino rats	100 mg/kg/b.w. (oral) subcutaneous.	1 ml/kg/b.w. (olive oil), -	AST, ALT, MDA↓, SOD, CAT↑	-	-	[61]
35	<i>Cinnamomum verum</i>	Wistar albino rats.	100 mg/kg/b.w. (oral)	1 ml/kg/b.w. (olive oil), i.p.	ALT, AST, ALP, γ-glutamyl transferase, LDH, TBARS↓	ALT, AST, ALP, γ-glutamyl transferase, LDH, TBARS↓	Flavonoids	[62]
36	<i>Cinnamomum zeylanicum L.</i>	Wistar rats	0.1 g/kg/oral)	0.5 ml/kg/b.w. (50% olive oil), 0.8 ml/kg (olive oil 1:1), i.p.	AST, ALT, MDA↓, SOD, CAT↑	AST, ALT, MDA↓, SOD, CAT↑	Flavonoids	[63]
37	<i>Citrus aurantium (essential oil)</i>	Sprague-Dawley rats	0.8 ml/kg/b.w. (i.p)	0.8 ml/kg (olive oil,50:50). Silymarin 50 mg/kg (i.p).	AST, ALT↓.	Limonene, α,β-pinene	[64]	
38	<i>Citrus limon(L.) Burm.F.</i>	Wistar rats	500 mg/kg/b.w. (oral)	1 ml/kg (olive oil,50:50). (oral).	ALT, AST, ALP, T, B, MDA↓, SOD, GSH, CAT, albumin↑	Coumarins, limonoids, flavonoids, eriocitrin, C-glycosyl flavones 6,8-di-C-β-glucosyl- diosmin	[65]	
39	<i>Clerodendrum volubile</i>	Wistar albino rats	500 mg/kg/bw.(oral)	1 ml/kg/b.w. (olive oil), i.p.	ALT, AST, ALP, LDH ↓, HDL, GSH, CAT, SOD, GPx↑	Phenols	[66]	
40	<i>Clitoria ternatea L.</i>	Wistar albino rats	300 mg/kg/bw (oral)	2.5 ml/kg/b.w. bw, (oral)	AST, ALT, ALP, Acid phosphatase(ACP), LDH, TB↓, TP↑	Flavonoid, terpe- noids, tannin, steroid, quercimetrin, rutin, scutellarein	[56]	
41	<i>Coriandrum sativum. L</i>	Wistar albino rats	300 mg/kg(i.p)	1 ml/kg/b.w. (liquid parafin, 1:1), oral. (i.p)	SGOT, SGPT, ALP↓, TB↑	Caffeic acid, quercetin, gallic acid, flavonoids, essential oil	[67]	
42	<i>Coriandrum sativum. L</i> (essential oil)	NMR mice	0.03 g/kg/b.w. (oral)	2 ml/kg/b.w. (olive oil, 1:2), i.p.	AST, ALT, LPx, XOD, Px↓, GSH, GSH-Px, Protein↑	Carvon	[55]	

Table 2 (continued)

s.no.	Botanical name	Animal model	Maximum extract dose/route of administration	CCL4 dose/route of administration	Standard drug administered/route of administration	Result.	Active components.	Reference
43	<i>Coriandrum sativum</i>	Wistar albino rats	200 mg/kg/b.w. (i.p)	1 ml/kg b.w. (1:1 olive oil), i.p.	Silymarin, 25 mg/kg/b.w., (i.p)	ALP AST, ALT, TP↑, TB, MDA↓, SOD, CAT, GPx↑	Caffeic acid, ferulic acid, isoquercitrin, rutin, quercetin 3-glucuronide, Quercetin, hyperin, quercetin-3-O-β-D-xyloside, quercetin-3-O-D-arabinose	[68]
44	<i>Cortex dictamni</i>	Sprague-Dawley rats	320 mg/kg/b.w. (oral)	2 ml/kg/b.w. i.p.	-	AST, ALT, ALP↓, SOD, CAT, GPx↑, GSH-Px, GSH↑, MDA↓	Limonoids, furoquinoline, flavonoids, fraxinellone	[69]
45	<i>Curcuma longa L.</i>	Sprague-Dawley rats	300 mg/kg/b.w. (intragastrically)	0.1 ml/kg/b.w. i.p.	Curcumin, 200 mg/kg/b.w., (intragastrically)	AST, ALT, TBARS↓, SOD, GPx↑.	Curcumin, memethoxy curcumin, bisdemethoxy curcumin	[70]
46	<i>Cytisus scoparius L.</i>	Wistar albino rats	500 mg/kg/b.w. (oral)	5 ml/kg(50% olive oil), i.p.	Silymarin, 25 mg/kg/b.w., (oral)	SGOT, SGPT, LDH↓, GSH, SOD, CAT, GPx, GRD, GST↑, TBARS↓	Rutin, quercetin, quer-citrin, isorhamnetin, kaempferol	[71]
47	<i>Dicoma anomala Sond</i>	Wistar rats; Rattus norvegicus	500 mg/kg/b.w. (oral)	1 ml/kg/b.w. (1:1 olive oil), i.p.	Silymarin, 100 mg/kg/b.w., (oral)	AST, ALT↓, SOD, CAT, GPx↑	Total flavonoid and phenol contents	[72]
48	<i>Dioscorea alata</i> peel	Wistar albino rats	433.42 mg/kg/b.w.	1 ml/kg (20% olive oil)	Silymarin, 200 mg/kg/b.w.	ALT, ALP AST, TBARS↓, SOD, CAT, GSH-Px↑, NO, TNF-α, TNF-Kb, iNOS, COX-2 expression	Hesperetin, quercetin, hesperidin	[73]
49	<i>Eclipta alba(L) Hassk</i>	Male albino rats	500 mg/kg/b.w. (oral)	2 ml/kg/b.w. (olive oil), i.p.	Silymarin, 50 mg/kg/b.w., (i.p).	ALT, AST, ALP, TB↓, TP↑	Flavonoids, luteolin, demethylivedolactone,wedelolactone	[74]
50	<i>Emblica officinalis</i> (Gaertn)	-	200 mg/kg/b.w.	1 ml/kg/b.w. (corn Oil), oral.	-	SGOT, SGPT, LDH, MDA↓, GSH, GST, GPx, GRx, TP↑, DNA synthesis↓	Quercetin, ascorbic acid, ellagic acid	[75]
51	<i>Entada purpurea</i>	Colony bred male Wistar rats	300 mg/kg/b.w. (oral)	2 ml/kg/b.w. (1:1 olive oil).	Silymarin, 50 mg/kg/b.w. (2% polyisorb-ate 80), (oral).	ALT, AST, ALP, TB↓, TP↑, LDH, MDA, Nitrate-nitrite, myeloperoxidase↓, SOD, CAT, GSH↑	Flavonoids	[76]
52	<i>Ephedra foliate Boiss</i>	Wistar rats	500 mg/kg/b.w.	0.125 ml/kg (liquid paraffin, 1:1), i.p.	Silymarin, 10 mg/kg, (oral).	SGOT, SGPT, ALP, TB	Flavonoids, tannins	[38]
53	<i>Euphorbia draconculloides</i> L.	Sprague-Dawley rats	400 mg/kg/b.w. (oral)	1 ml/kg/b.w. (30% olive oil), i.p.	Silymarin 50 mg/kg/b.w.	AST, ALT, ALP↓, CAT, Peroxi-dase, SOD, GST, GSH↑, Lipid peroxides, TBARS, nitrite, hydrogen peroxide, DNA damage↓	Catechin, rutin, caffeo-acid, mircetin, cou-marins, flavonoids	[5]

Table 2 (continued)

s.no.	Botanical name	Animal model	Maximum extract dose/route of administration	CCL4 dose/route of administration	Standard drug administered/route of administration	Result	Active components.	Reference
54	<i>Fagonia schweinfurthii</i> (<i>Haddidi</i>)	Wistar albino rats	400 mg/kg/b.w. (oral)	1 ml/kg/b.w., i.p.	Silymarin 100 mg/kg/b.w., (oral)	ALT, AST, ALP, TB _↓ , MDA _↓ , SOD, CAT, GSH _↑	Flavonoids, phenolic compounds, quinines and, coumarin	[77]
55	<i>Ficus carica</i> Linn	Wistar rats	100 mg/kg/b.w. (oral)	1 ml/kg/b.w/v olive oil, i.p.	—	SGOT, SGPT, T.B. ↓	Psoralen, bergapten, xantho toxin, calotropin acetate, lupeol	[78]
56	<i>Flemingia macrophylla</i>	Male SD rats	1.0 g/kg/b.w. (oral)	15 ml/kg/b.w. (20% olive oil), i.p.	Silymarin, 25 mg/kg/b.w. in carboxy methyl cellulose.	ALT, AST, MDA _↓ , SOD, CAT, GSH-Px, GSH _↑ , NO, TNF- α , IL-1 β _↓ .	Genistein, lupeol, rutin, flavonoids, Isoflavones	[79]
57	<i>Ginkgo biloba</i>	Sprague-Dawley rats	150 mg/kg/b.w. (oral)	1 ml/kg/b.w. (1:1 liquid paraffin).	—	ALP, ALT, AST, MDA _↓ , TP, HDL-C, GSH _↑	Kaempferol, quercetin, isorhamnetin, diterpenes lactones	[80]
58	<i>Glyphaea brevis</i>	Swiss albino mice	490 mg/kg/b.w. (oral)	2 ml/kg/b.w. (liquid paraffin) i.p.	Silymarin, 100 mg/kg (oral)	GSH, CAT, SOD _↑ , TBARS, ALT, AST, ALP, T-chol, LDL, TG _↓ , TP _↑	Flavonoids	[81]
59	<i>Graptophetalum paraguayense</i> E. Walter	Sprague-Dawley rats	300 mg/kg/b.w. (oral)	0.5 ml/kg/b.w. (1:4 olive oil) oral.	Silymarin, 200 mg/kg/b.w., (oral)	AST, ALT, MDA _↓ , GSH, SOD, GR, SOD, CAT _↑ , TNF- α _↓	Gallic acid, genistin, daidzin, quercetin	[82]
60	<i>Hibiscus sabdariffa</i> . L.	Wistar rats	500 mg/kg/b.w.	0.125 ml/kg (liquid paraffin, 1:1), i.p.	Silymarin, 10 mg/kg (oral)	—	—	[38]
61	<i>Hippophaea rhamnoides</i> L	C57BL/6 mice	200 mg/kg/b.w. (oral)	5 ml/kg/b.w. (20% in peanut oil), i.p.	—	ALT, AST, TB _↓ , PALB, SOD, GSH-Px, GSH _↑ , MDA, TNF- α , IL-1 β , iNOS, NO, TLR4, p38MAPK, p-ERK, p-JNK, NF- κ B _↓	Isorhamnetin, quercetin, chlorogenic acid, myricetin, kaempferol, catechins	[83]
62	<i>Indigofera oblongifolia</i>	Wistar albino rats	300 mg/kg/b.w. (oral)	1 ml/kg/b.w. (30% olive oil), i.p.	—	ALT, AST, ALP, TB _↓ , GSH, SOD, CAT, GPx _↑	Flavonoids, coumarins, indirubin	[84]
63	<i>Launaea procumbens</i>	Sprague-Dawley rats	200 mg/kg/b.w. (oral)	3 ml/kg/b.w. (30% olive oil), i.p.	Silymarin 100 mg/kg/b.w., (oral)	AST, ALT, ALP, LDH _↓ , GST, GSH, GSH, CAT, POD, SOD, GSH-Px _↑	Salicylic acid, vanillic acid, synergic acid, 2-methyl-resorcinol, and gallic acid	[85]
64	<i>Lawsonia inermis</i> L (Henna)	Albino rats	200 mg/kg/b.w. (oral)	2 ml/kg/b.w (1:1 olive oil).	Silymarin, 25 mg/kg/b.w. (oral).	ALT, AST, ALP, TB _↓ , TP _↑	Flavonoids	[86]
65	<i>Lawsonia inermis</i> Linn	Wistar albino rats	400 mg/kg/b.w. (i.p.)	1.25 ml/kg(1:1 liquid paraffin), i.p.	Silymarin, 100 mg/kg/b.w. (i.p.)	SGOT, SGPT, MDA _↓ , TP, GSH _↑	Flavonoids	[87]
66	<i>Leucas cephalotes</i> Linn	Wistar albino rats	200 mg/kg/b.w. (liquid paraffin) (i.p.)	1.25 mg/kg (1:1 liquid paraffin), i.p.	Silymarin, 200 mg/kg (i.p.)	SGOT, SGPT, Alkaline phosphatase (ALKP), TB _↓ , TP, TC, ↑.	Flavonoids	[88]

Table 2 (continued)

s.no.	Botanical name	Animal model	Maximum extract dose/route of administration	CCL4 dose/route of administration	Standard drug administered/route of administration	Result.	Active components.	Reference
67	<i>Lobularia maritima</i>	Mice	500 mg/kg/b.w. (i.p)	1 ml/kg/b.w. (1:1 olive oil), i.p.	—	Silymarin, 25 mg/kg/b.w. (oral)	p-coumaric acid	[7]
68	<i>Luffa acutangula</i> (Var) <i>amara</i>	Colony bred strain of Wistar rats	600 mg/kg/b.w. (oral)	1 ml/kg/b.w. oral	Silymarin, 50 mg/kg/b.w. (oral)	IL-1 β , IL-6L, SOD, CAT, GPx \uparrow	Flavonoids	[89]
69	<i>Lygodium flexuosum</i> (L.) Sw	Wistar rats	200 mg/kg/b.w.	150 μ l /100 g (1:1 corn oil)	Silymarin, 50 mg/kg	SGOT, SGPT, ALP, TC, TB \downarrow , TP, GPx, GST, GSH, SOD, CAT \uparrow , LPO \downarrow , Vit E, Vit C, \uparrow	B-sitosterol, stigmasterol, kaempferol, tectoquinone	[90]
70	<i>Madhuca indica</i> Syn	Wistar rats	400 mg/kg/b.w (oral)	2 ml/kg/b.w. (olive oil), (i.p)	Silymarin, 100 mg/kg/b.w.	AST, ALT, MDA, ROS, TNF- α	Flavonoids	[91]
71	<i>Madhuca indica</i> Syn	Wistar rats	300 mg/kg/b.w.	0.5 ml/kg/b.w., i.p.	Silymarin, 100 mg/kg/b.w.	SGOT, SGPT, ALP, TB \downarrow	Flavonoids	[92]
72	<i>Mahonia aquifolium</i>	Wistar albino rats	500 mg/kg/b.w (oral)	1 ml/kg/b.w. (50% olive oil), i.p.	Silymarin, 200 mg/kg/b.w. (oral)	ALT, AST, MDA \downarrow , SOD, GPx, GR \uparrow , TNF- α , NO \downarrow	Berberine, palmatine, jatrorrhizine	[3]
73	<i>Mallotus philippensis</i> Muell-Arg	Wistar albino rats	200 mg/kg/b.w (oral)	600 mg/kg/ml, oral	Silymarin, 25 mg/kg/b.w. (oral)	SGPT, SGOT, ALP, TBL, TP, CAT, SOD \uparrow , LPO \downarrow	Flavonoids, phenols, isocoumarins, bergenin	[93]
74	<i>Memnonia tuberosa</i> Cogn	Wistar rats	400 mg/kg/b.w (oral)	2 ml/kg/b.w(1:1 liquid paraffin), subcutaneous	Silymarin, 100 mg/kg, (oral)	SGOT, ALP, TB, Cholesterol, TAG, MDA \downarrow , GSH \uparrow	Vitamin C, saponins, triterpenoids	[94]
75	<i>Mentha piperita</i> L.	Wistar rats	40 mg/kg/b.w (oral)	1 ml/kg (olive oil), i.p.	Silymarin, 50 mg/kg/b.w. (oral)	ALT, AST, ALP, LDH, γ -GT \downarrow , SOD, CAT, GPx \uparrow , TBARS \downarrow	Spathulenol, cadinene, caryophyllene, caryophyllene oxide	[95]
76	<i>Mentha arvensis</i> Linn	Albino wistar rats	375 mg/kg/b.w (oral)	0.5 ml/kg/b.w, i.p.	Silymarin, 100 mg/kg/b.w. (oral)	SGPT, SGOT, ALP, TB \downarrow	Luteolin, mentholide, rutin, hesperidin, phenolic acid quercetin, isorhamphol	[96]
77	<i>Mimosa pudica</i> 2009	Wistar albino rats	200 mg/kg/b.w (oral)	1.25 ml/kg/b.w. (1:1 liquid paraffin), i.p.	Silymarin, 100 mg/kg/b.w.	SPGT, SGOT, ALP, TBL, T chole \downarrow , TP, albumin \uparrow	Flavonoids, alkaloids, glycosides	[97]
78	<i>Mimosa pudica</i> Linn	Wistar albino rats	400 mg/kg/b.w. (oral)	1 ml/kg/b.w (1 : 2 liquid paraffin), subcutaneous.	Silymarin, 10 mg/kg/b.w. (oral)	SGOT, ALP, TB, SGPT \downarrow	Flavonoids, phenols, gallic acid	[98]
79	<i>Momordica dioica Roxb</i>	Wistar albino rats	200 mg/kg/b.w. (oral)	2 ml/kg/bw(1:1 liquid paraffin).	Silymarin, 5 mg/kg/b.w (oral)	AST, ALT, ALP, TB, MDA \downarrow , SOD, CAT, GSH \uparrow , Hydroperoxides \downarrow	Flavonoids, phenolic compounds	[99]
80	<i>Nerium oleander</i> Linn	Wistar rats	400 mg/kg/b.w. (oral)	1 ml/kg/bw(1:1 olive oil), i.p.	Silymarin, 100 mg/kg/b.w. (oral).	AST, ALT, ALP, TB, MDA \downarrow , SOD \uparrow	Oleandrin, Oleanolic acid	[100]
81	<i>Nicotiana plumbaginifolia</i> L.	Male chicks	200 mg/kg/b.w. (oral)	1 ml/kg/bw (30% olive oil), i.p.	Silymarin, 100 mg/kg/b.w. (gavage).	CAT, Peroxidase, SOD, GP-X, GRS \uparrow , TBARS, LDH, TAG, T.Chol, LDL \downarrow , HDL \uparrow	Rutin, chlorogenic acid, quercetin	[101]

Table 2 (continued)

s.no.	Botanical name	Animal model	Maximum extract dose/route of administration	CCL4 dose/route of administration	Standard drug administered/route of administration	Result.	Active components.	Reference
82	<i>Nymphaea alba</i> L.	Wistar albino rats	200 mg/kg/b.w. (oral)	0.5 ml/kg/b.w, i.p.	Silymarin, 100 mg/kg/b.w(oral).	MDA↓, GSH↑, CAT, SOD, TAC↑, TNF-α, Caspase-3↓	Phenols, flavonoids, quercentin, ellagic acid, gallic acid, kaempferol [6]	
83	<i>Olea europaea</i> L.	Sprague-Dawley rats	80 mg/kg/b.w. (oral)	0.2 ml/kg/b.w, i.p.	—	ALP, AST, ALP↓, CAT, SOD↑	Caffeic acid, diosmetin, verbascoside, oleuropein, luteolin 7-O-glucoside, rutin, leuteolin 4'-O-glycoside, P-coumaric acid, vanillin [102]	
84	<i>Origanum vulgare</i>	Wistar albino rats	150 mg/kg/b.w. (oral)	2 ml/kg/b.w (1:1 olive oil)	—	ALT, ALP, AST↓, LPO, GST, CAT, SOD, GPx, GR, GSH↑	Carvacrol, thymol [103]	
85	<i>Persea Americana</i> mill	Wistar albino rats	200 mg/kg/day	3 ml/kg(1:1 olive oil) subcutaneous	reducodyn®, 100 mg/kg/day	ALT, AST, ALP, TB↓, CAT, SOD, GPx, GST↑, Protein carbonyl↓	Flavonoids [104]	
86	<i>Phyllanthus niruri</i>	Wistar rats	100 mg/kg/b.w. (oral)	1 ml/kg/b.w(50% in corn oil), i.p.	Silymarin, 1mg/ml, (i.p.)	AST, ALT, ALP, LDH, T-cho, TB↓, TP↑, TNF-α, NF-κB, IL-6, IL-8, IL-10↓, GR, GP↑, MDA↓, GSH↑, ROS↓.	Quercetin, gallic acid, corilagin, isoconifagin, hamnoside, brevifolin carboxylic acid [105]	
87	<i>Physalis peruviana</i>	Wistar albino rats	500 mg/kg/b.w (oral)	0.5 ml/kg/bw (olive oil), i.p.	legation®, 100 mg/kg/b.w. (oral).	MDA↓, SOD↑, NO, AST↓, ALT↑, ALP↓, TB, TP↓	Flavonoids, lupeol, ursolic acid [106]	
88	<i>Pleioignytum timorense</i> (DC) Leenh	Sprague-Dawley rats	300 mg/kg/b.w.	0.5 ml/kg(10% olive oil)	Silymarin 50 mg/kg/b.w.	AST, ALT↓, TAC↑	Catechin, gallic acid, kaempferol, quercetin, β-sitosterol, lupeol [107]	
89	<i>Pleurorus ostreatus</i>	Wistar albino rats	200 mg/kg/b.w. (i.p)	2 ml/kg/b.w (olive oil), i.p.	—	SGOT, SGPT, ALP, MDA↓, GSH, CAT, SOD, GPx↑	— [108]	
90	<i>Polygonum cuspidatum</i> Sieb et Zucc	Male ICR mice	100 mg/kg/day (oral)	50 µl/kg (olive oil) i.p.	Bifendate, 150 mg/kg/b.w. (oral)	AST, ALT, MDA, TNF-α, IL-1β, COX-2, iNOS, NF-κB↓, SOD, GST, GSH, CAT, GPx, TGF-β1↑	Polydatin, resveratrol, quercentin, emodin, citreorosein [4]	
91	<i>Premna esculetina Roxb</i>	Long-Evans rats (<i>Rattus norvegicus</i>)	400 mg/kg/day (oral)	1 ml/kg/bw (1:1 olive oil), i.p.	Silymarin, 100 mg/kg/day, (oral)	SGPT, SGOT, SALP↓, TP, ALB↑	Phenols, tannins, flavonoids [109]	
92	<i>Raphanus sativus</i>	Albino rats	300 mg/kg/b.w. (oral)	1 ml/kg/bw (1:1 olive oil)	Silymarin, 50 mg/kg/b.w. (oral)	AST, ALT, ALP, TB↓, CAT, GSH↑, MDA↓	Flavonoids, polyphenols [110]	
93	<i>Rourea indica planch</i>	Wistar albino rats	500 mg/kg/b.w. (oral)	2 ml/kg/b.w, i.p.	Legalon, 50 mg/kg/b.w. (oral)	AST, ALT, TB↓, CAT, SOD, GPx, GSH↑, TBARS↓	Hyperin, quercentin-3-O-β-D-xyloside, quercentin-3-D-α-arabinofuranoside, quercentin [111]	

Table 2 (continued)

s.no.	Botanical name	Animal model.	Maximum extract dose/route of administration	CCL4 dose/route of administration	Standard drug administered/route of administration	Result.	Active components.	Reference
94	<i>Rubia cordifolia</i> Linn	Sprague-Dawley rats	200 mg/kg/b.w (oral)	0.1 ml/kg/b.w, i.p.	Silymarin, 100 mg/kg/b.w. (oral).	SGOT, SGPT, SAKP, γ-GT↓, GST, GR, GSH↑, MDA↓	Rubiadin	[112]
95	<i>Rumex vesicarius</i> L	Wistar albino rats	200 mg/kg/ b.w (oral)	1.5 ml/kg/ b.w (1% tween 80) i.p.	Silymarin 50 mg/kg/b.w.	SGOT, SGPT, ALP↓, TP↑, TBL, CAT, SOD↑, MDA↓	Phenols, flavonoids	[113]
96	<i>Semen celosia Cristatae</i> L	Kunming mice	4.0 mg/kg/b.w (oral)	0.1% (edible oil), i.p.	Bifendate	AST, ALT, ALP, MDA↓, GSH-Px, CAT, SOD↑	Semenoside	[114]
97	<i>Solanum trilobatum</i> Linn	Wistar Albino rats	250 mg/ kg/b.w (i.P)	1 ml/kg/bw(30% olive oil), i.p.	–	ALT, AST, ALP, LDH↓, TP, GSH GPx, CAT, SOD↑, Lipid peroxide↓	Sobatum, solasodine, β-solanamine, solaine	[115]
98	<i>Solanum xantholarpum</i>	Sprague-Dawley rats	400 mg/kg/bw (oral)	1 ml/kg (1:1 liquid paraffin)	Silymarin, 100 mg/kg/b.w. (oral)	AST, ALT, ALP, TB, MDA↓, CAT, GSH, SOD↑	Flavonoids, quercetin	[116]
99	<i>Spondias mombim</i>	Wistar rats	1000 mg/kg/b.w (oral)	2 ml/kg/b.w. (1:1 liquid paraffin)	Silymarin, 100 mg/kg/b.w. (oral)	ALT, AST, ALP, TBL, GSH, CAT, SOD↑, TBARS↓	Flavonoids, phenols	[117]
100	<i>Stachys pilifera</i> Benth	Wistar rats	400 mg/kg/day (oral)	1 ml/kg/b.w. (50% olive oil)	–	AST, ALT, ALP, MDA↓, TP, T.B↑,	Flavonoids, phenylethanoid glycosides, diterpenes, terpenoids	[118]
101	<i>Vitis thunbergii</i> var	Male SD rats	400 mg/kg/b.w.	1.5 ml/kg/b.w. (20% olive oil) i.p.	Silymarin, 200 mg/kg/b.w. in carboxy methylcellulose	ALT, AST, MDA↓, SOD, CAT, GPx, GSH↑, TNF-α, IL-1β, NO, iNOS, COX-2↓	Resveratrol derivatives, polyphenols compounds, quercetin, oligostibenes	[119]
102	<i>Xylaria nigripes</i> (Koltz) Sacc	ICR mice	100 mg/kg/b.w. (intragastrically)	2 ml/kg/b.w. (40% olive oil). Subcutaneously	Silymarin, 100 mg/kg/b.w., (intragastrically)	SGOT, SGPT, TBARS↓, SOD, CAT, GPx, ↑	Epicatechin, P-coumaric acid, catechin	[120]
103	<i>Zingiber officinale</i> (Roscoe) rhizome (Ginger)	Wistar rats	400 mg/kg/b.w. (oral)	0.7 ml/kg/b.w. (1:1, olive oil)	Livolin fort®, 5.2 mg/kg/b.w. (oral)	AST, ALT, ALP↓, TP, GSH, CAT↑,	Flavonoids, 6-gingerol, shogaols	[121]
104	<i>Ziziphus jujube</i>	Male ICR mice	200 mg/kg/bw. (intragastrically)	2 ml/kg/bw. (40% olive oil), subcutaneously	Bifendate, 7.5 mg/kg/b.w., (intragastrically)	ALT, AST, MDA↓, SOD, CAT, GSH-Px, GSH↑	Flavonoids	[122]

↓ decrease in effect/activity; ↑ increase in effect/activity

formation of CCl_3^* and CHCl_2^* and $\text{CCl}_3\text{-OO}^*$ radicals, lipid peroxidation, membrane damage, the severe derailment of intracellular Ca^{2+} sequestration, apoptosis, and fibrosis [10, 30, 31].

Traditional plants with anti-hepatotoxic potential

In this review, numerous experimental studies on the medicinal plants effectiveness to ameliorate CCl_4 -induced hepatotoxicity in animal models were presented. The botanical names, ethnopharmacological and pharmacological uses of plants traditionally used to treat liver-related diseases were presented in Table 1. The comprehensive details on in vivo studies of medicinal plants with hepatoprotection against CCl_4 -induced hepatotoxicity alongside the active phytochemicals and their probable mechanisms of action are presented in Table 2.

Discussion

For about three decades, extracts from different natural products have been identified to be hepatoprotective at varied doses against CCl_4 -induced toxicity by reducing oxidative stress on liver enzymes. The findings from this review show that only few studies tested these natural products on hepatic cell lines (Table 2). Without separating the whole extract to identify the active components, a large number of hepatoprotective products will increase without corresponding clinical relativity [123]. There is an urgent need to study individual components of the plant extract especially in experimental animal models. The major drawback of herbal medicine is its potential hepatotoxicity in man which could cause acute to chronic liver injury with underlining mechanism of toxicity not clearly understood due to factors such as the synergistic and multi-organ targeted nature of the various components [124–127].

The protection provided by herbal plants against CCl_4 -induced hepatotoxicity is basically due to the inhibitory nature of the phytochemicals present in them [70, 101]. These phytochemicals are able to inhibit the microsomal enzymes to restrict the generation of free radicals and stop lipid peroxidation through its antioxidant ability [66]. They can also enhance the regeneration of liver cells, radical scavenging, and stimulation of the anti-inflammatory ability of the liver cells against the inflammation induced by CCl_4 [102].

The treatment of the animal models with these herbal extracts showed beneficial effects through several biochemical and histological results. From the results in Table 2, it is clear that these plants extract downregulated serum liver marker enzymes like aspartate transaminase (AST), alanine transaminase (ALT), alkaline phosphatase (ALP), total bilirubin, and malondialdehyde (MDA) while

upregulating the activity of antioxidant enzymes and total protein. The medicinal plants also downregulated the inflammatory markers expression in the hepatic cells. Some of these reported studies confirmed the hepatoprotective effectiveness of these medicinal plant products through histological reports [43, 54]. This review also reported numerous phytochemicals with possible hepatoprotective potentials ranging from flavonoids (quercetin, kaempferol), phenols, saponins, vitamin C, caffeic acid, etc. This review presented a number of plant species with ethnopharmacological relevance in the treatment of liver injury and their medicinal/pharmacological uses from literature.

Conclusion

We, therefore, conclude that there are a variety of phytochemicals in plant products with hepatoprotective activity against CCl_4 -induced toxicity by downregulation of liver marker enzymes, and activation of antioxidative capacity of the liver cells that leads to the restoration of the liver architecture.

Future perspectives

There is need to validate the efficacy of some of the reported active components which can be likely candidate for therapeutic purposes. Research should move from whole plant extract experiment to isolation of bioactive components and testing the extract on culture cell lines.

Abbreviations

ALT: Alanine transaminase; AST: Aspartate transaminase; ALP: Alkaline phosphatase; γ -GT: Gamma glutamyltransferase; LDH: Lactate dehydrogenase; MDA: Malondialdehyde; GSH: Glutathione; GPx: Glutathione peroxidase; CAT: Catalase; SOD: Superoxide dismutase; POD: Peroxidase; GST: Glutathione S-transferase; GST α : Glutathione S-transferase alpha; GR: Glutathione reductase; TBARS: Thiobarbituric acid reactive substance; NO: Nitric oxide; H_2O_2 : Hydrogen peroxide; TNF- α : Tumor necrosis factor alpha; NF- κ B: Nuclear factor kappa B; iNOS: Inducible nitric oxide synthase; COX-2: Cyclo oxygenase-2; IL-1 β : Interlukin-1 beta; NrF-2: Nuclear factor erythroid-2-related factor 2; TGF- β (1): Hepatic growth factor-beta 1; IL-6: Interleukin-6; IL-8: Interleukin-8; IL-10: Interleukin-10; HO-1: Heme oxygenase-1; NP-SH: Nonprotein sulfhydryls; NQO1: Quinine oxidoreductase; TLR4: Hepatic toll-like receptor 4; P38MAPK: P38 mitogen-activated protein kinase; p-ERK: Extracellular signal-regulated kinase; p-JNK: C-jun N-terminal kinase; CYT: Cytochrome; DTdiaphorase: A phase II enzyme; T-cho: Total cholesterol; TG: Triglycerides; LDL: Low-density lipoprotein; TAG: Triacylglycerol; HDL: High-density lipoprotein; TP: Total protein; TB: Total bilirubin; XOD: Xanthine oxidase; Vit. A: Vitamin A; Vit. E: Vitamin E; Vit. C: Vitamin C; CNS: Central nervous system.

Acknowledgements

Not applicable.

Authors' contributions

CEU conceived the idea and wrote the initial draft. SMS did the literature search and data collection. Both authors proof read the final manuscript.

Funding

Not applicable.

Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Declarations

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declared no competing interests.

Received: 1 December 2020 Accepted: 23 November 2021

Published online: 09 December 2021

References

- Zhang X, Feng J, Su S, Huang L (2020) Hepatoprotective effects of *Camellia nitissima* aqueous ethanol extract CCl₄-induced acute liver injury in SD rats related to Nrf2 and NF-κB signaling. *Pharm Biol* 5(1):239–246
- Wang S, Luan JJ, Lv XW (2019) Inhibition of endoplasmic reticulum stress attenuated ethanol-induced exosomal miR-122 and acute liver injury in mice. *Alcohol Alcohol* 54:465–471
- Chao J, Lee M-S, Amagaya S, Liao J-W, Ho L-K, Peng W-H (2009) Hepatoprotective effect of *Shidagronlo* on acute liver injury induced by carbon tetrachloride. *Am J Chin Med* 37(6):1085–1097
- Zhang H, Yu C-H, Jiang Y-P, Peng C, He K, Tang J-Y, Xin H-L (2012) Protective effects of polydatin from *Polygonum cuspidatum* against carbon tetrachloride-induced liver injury in mice. *PLoS One* 7(9):e46574. <https://doi.org/10.1371/journal.pone.0046574>
- Batool R, Khan MR, Majid M (2017) *Euphorbia dracunculoides* L. abrogates carbon tetrachloride induced liver and DNA damage in rats. *BMC Complement Altern Med* 17:23. <https://doi.org/10.1186/s12906-017-0174-x>
- Bakr RO, El-Naa MM, Zaghlou SS, Omar MM (2017) Profile of bioactive compounds in *Nymphaea alba* L. leaves growing in Egypt: hepatoprotective, antioxidant and anti-inflammatory activity. *BMC Complement Altern Med* 17:52. <https://doi.org/10.1186/s12906-017-1562>
- Hsouna AB, Dhibi S, Dhifi W, Saad RB, Brini F, Hfaidh N, da Silva Almeida JRG, Mnif W (2020) *Lobularia maritima* leave extract, a neutraceutical agent with antioxidant activity, protects against CCl₄-induced liver injury in mice. *Drug Chem Toxicol*. <https://doi.org/10.1080/01480545.2020.1742730>
- Jones I (1983) Chloroform anaesthesia in Liverpool. *Anaesthesia* 38:578–580
- Agency for Toxic Substances and Disease Registry (ATSDR) (2005) Toxicological profile for carbon tetrachloride. U.S. Department of Health and Human Services, Public Health Service, Atlanta
- Clawson GA (1989) Mechanism of carbon tetrachloride toxicity. *Pathol Immunopathol Res* 8:104–112
- Recknagel R, Lombardi B (1961) Studies of biochemical changes in subcellular particles of rat liver and their relationship to a new hypothesis regarding the pathogenesis of carbon tetrachloride fat accumulation. *J Biol Chem* 236:564–569
- Judah J (1969) Biochemical disturbances in liver injury. *Br Med Bull* 25:274–277
- de Vries J (1983) Induction and prevention of biochemical disturbances in hepatic necrosis. *Trends Pharmacol Sci* 4:393–394
- Smuckler E, Iseri O, Bendit E (1962) An intracellular defect in protein synthesis induced by carbon tetrachloride. *J Exp Med* 116:55–72
- Moore L, Chen J, Knapp H, Landon E (1975) Energy-dependent calcium sequestration activity in rat liver microsomes. *J Biol Chem* 250:4562–4568
- Fulceri R, Benedetti A, Comporti M (1984) On the mechanisms of the inhibition of calcium sequestering activity of liver microsomes in bromothrichloromethane intoxication. *Res Commun Chem Pathol Pharmacol* 46:235–243
- Christie G, Judah J (1954) Mechanism of action of CCl₄ on liver cells. *Proc R Soc Lond Ser 142:241–257*
- De Groot H, Littauer A, Hugo-Wissmann D, Wissmann P, Noll T (1988) Lipid peroxidation and cell viability in isolated hepatocytes in a redesigned oxystat system: Evaluation of the hypothesis that lipid peroxidation, preferentially induced at low oxygen partial pressure, is decisive for CCl₄ liver cell injury. *Arch Biochem Biophys* 264:591–599
- Masuda Y, Nakamura Y (1990) Effects of oxygen deficiency and calcium omission on carbon tetrachloride hepatotoxicity in isolated perfused livers from phenobarbital-pretreated rats. *Biochem Pharmacol* 40:1865–1876
- Kiezcka H, Kappus H (1980) Oxygen dependence of CCl₄-induced lipid peroxidation *in vitro* and *in vivo*. *Toxicol Lett* 5:191–196
- Dianzani MU, Poli G (1985) Lipid peroxidation and haloalkylation in CCl₄-induced liver injury. In: Poli G, Cheeseman KH, Dianzani MU, Slater TF (eds) *Free Radicals in Liver Injury*. IRL Press, Oxford
- Dianzani MU (1984) Lipid peroxidation and haloalkylation: Two distinct mechanisms for CCl₄-induced liver damage. In: Calandra S, Carulli N, Salvio G (eds) *Liver and Lipid Metabolism*. Excerpta Medica, Elsevier, Amsterdam, New York, Oxford
- Marinari UM, Pronzato MA, Cottalasso D, Zicca-Cadoni A, Nanni G, Poli G, Chiarpotto E, Albano E, Biasi F, Dianzani MU (1985) CCl₄-induced early functional impairments of rat liver Golgi apparatus. In: Poli G, Cheeseman KH, Dianzani MU, Slater TF (eds) *Free Radicals in Liver Injury*. IRL Press, Oxford
- Cheeseman KH, Albano EF, Tomasi A, Slater TF (1985) Biochemical studies on the metabolic activation of halogenated alkanes. *Environ Health Perspect* 64:85–101
- Boll M, Weber LWD, Becker E, Stampfli A (2001a) Mechanism of carbon tetrachloride-induced hepatotoxicity. Hepatocellular damage by reactive carbon tetrachloride metabolites. *Z Naturforsch C. J Biosci* 56(7-8):649–659
- Ozaki M, Masuda Y (1993) Carbon tetrachloride-induced cell death in perfused livers from phenobarbital-pretreated rats under hypoxic conditions and various ionic milieu. Further evidence for calcium-dependent irreversible changes. *Biochem Pharmacol* 46:2039–2049
- Liu SL, Degli Esposti S, Yao T, Diehl AM, Zern MA (1995) Vitamin E therapy of acute CCl₄-induced hepatic injury in mice is associated with inhibition of nuclear factor Kappa B binding. *Hepatology* 22:1474–1481
- Czaja MJ, Xu J, Alt E (1995) Prevention of carbon tetrachloride-induced rat liver injury by soluble tumor necrosis factor receptor. *Gastroenterol* 108:1849–1854
- Kull FC, Cuatrecasas P (1981) Possible measurements of internalization in the mechanism of *in vitro* cytotoxicity of tumor necrosis serum. *Cancer Res* 41:4885–4890
- Boll M, Weber LWD, Becker E, Stampfli A (2001b) Pathogenesis of carbon tetrachloride-induced hepatocyte injury. Bioactivation of CCl₄ by cytochrome P₄₅₀ and effects on lipid homeostasis. *Z Naturforsch* 56c:111–121
- Boll M, Weber LWD, Becker E, Stampfli A (2001c) Hepatocyte damage induced by carbon tetrachloride. Inhibited lipoprotein secretion and altered lipoprotein composition. *Z Naturforsch* 56c:283–290
- Ai G, Liu Q, Hua W, Huang Z, Wang D (2013) Hepatoprotective evaluation of the total flavonoids extracted from flowers of *Abelmoschus manihot* (L) Medic: *In vivo* and *in vivo* studies. *J Ethnopharmacol* 146:794–802
- Arbab AH, Parvez MK, Al-Dosari MS, Al-Rehaily AJ, Al-Sohaibani M, Zaroug EE, Alsaïd MS, Rafatullah S (2015) Hepatoprotective and antiviral efficacy of *Acacia mellifera* leaves fractions against Hepatitis B virus. *Biomed Res Int*. <https://doi.org/10.1155/2015/929131>
- Singh R, Rao HS (2008) Hepatoprotective effect of the pulp/seed of *Aegle marmelos correa ex Roxb* against carbon tetrachloride induced liver damage in rats. *Inter J Green Pharm*:232–234
- Rathee D, Kamboj A, Sidhu S (2018) Augmentation of hepatoprotective potential of *Aegle marmelos* in combination with piperine in carbon tetrachloride model in wistar rats. *Chem Cent J* 12:94. <https://doi.org/10.1186/s3065-018-0463-9>

36. Dhruve P, Nauman M, Kale KR, Singh RP (2020) A novel hepatoprotective activity of *Alangium salvifolium* in mouse model. *Drug Chem Toxicol*. <https://doi.org/10.1080/01480545.2020.1733593>
37. Gargoun HM, Muftah SS, Shalmani SA, Mohammed HA, Alzoki AN, Debani AH, Fituri OA, Shari FE, Barassi IE, Meghil SE, Abdellatif AW (2013) Phytochemical screening and investigation of the effect of *Alhagi maurorum* (camel thorn) on carbon tetrachloride, acetaminophen and Adriamycin induced toxicity in experimental animals. *J Sci Innov Res* 2(6):1026–1033
38. Salah IA, Adnan JA, Abdulmalik MA, Maged SA (2008) Evaluation of hepatoprotective effect of *Ephedra foliata*, *Alhagi maurorum*, *Capsella bursa-pastoris* and *Hibiscus sabdariffa* against experimentally induced liver injury in rats. *Nat Prod Sci* 14(2):95–98
39. Naji KM, Al-Shaibani ES, Alhadi FA, Al-Soudi SA, D'souza MR (2017) Hepatoprotective and antioxidant effects of single clove garlic against CCl_4 -induced hepatic damage in rabbits. *BMC Complement Altern Med* 17:411. <https://doi.org/10.1186/s12906-017-1916-8>
40. Zeashan H, Amresh G, Singh S, Rao CV (2008) Hepatotoxicity activity of *Amaranthus spinosus* in experimental animals. *Food Chem Toxicol* 46:3417–3421
41. Jain S, Dixit VK, Malviya N, Ambawatia V (2009) Antioxidant and hepatoprotective activity of ethanol and aqueous extracts of *Amorphophallus cmanulatus Roxb.* tubers. *Acta Pol Pharm Drug Res* 66(4):423–428
42. Sourabie TS, Ouedraogo N, Sawadogo WR, Yougbare N, Nikiema JB, Guissou IP, Nacoulma OG (2012) Evaluation of the anti-icterus effect of crude powdered leaf of *Argemone Mexicana L.* (*Papaveraceae*) against CCl_4 -induced liver injury in rats. *IJPSR* 3(10):491–496
43. Wang J-H, Choi M-K, Shin J-W, Hwang S-Y, Son C-G (2012) Antifibrotic effects of *Artemisia capillaris* and *Artemisia iwayomogi* in a carbon tetrachloride-induced chronic hepatic fibrosis animal model. *J Ethnopharmacol* 140:179–185
44. Surenda HB, Apana R (2007) Hepatoprotective properties of *Bauhinia variegata* bark extract. *Yakugaku Zasshi* 127(9):1503–1507
45. Gilani AH, Janbaz KH, Shah BH (1998) Esculetin prevents liver damage induced by paracetamol and CCl_4 . *Pharm Res* 37(1):31–35
46. Enas JK (2014) Phytochemicals investigation and hepato-protective studies of Iraqi *Bryonia* (Family *Cucurbitaceae*). *IJPSR* 6(4):187–190
47. Akindele AL, Ezenwanebe KO, Anunobi CC, Adeyemi OO (2010) Hepatoprotective and *in vivo* antioxidant effects of *Byrsocarpus co-Cneus Schum.* and *Thonn* (*Connaraceae*). *J Ethnopharmacol* 129:40–52
48. Singh S, Mehta A, Mehta P (2011) Hepatoprotective activity of *Cajanus cajan* against carbon tetrachloride induced liver damage. *IJPSR* 3(sup 2):146–147
49. Lodhi G, Singh HK, Pant KK, Hussain Z (2009) Hepatoprotective effects of *Calotropis gigantean* extract against carbon tetrachloride induced liver injury in rats. *Acta Pharm* 59:89–96
50. Josi YM, Kadam VJ, Patil YV, Kaldhone PR (2009) Investigation of hepatoprotective activity of aerial parts of *Canna indica L.* on carbon tetrachloride treated rats. *J Pharm Res* 2(12):1879–1882
51. Nasrin A, Iran R, Amir M (2007) Hepatoprotective activity of *Capparis spinosa* root bark against CCl_4 induced hepatic damage in mice. *Iranian J Pharm Res* 6(4):285–290
52. Saheen S, Khan MR, Khan RA (2011) Hepatoprotective effects of methanol extract of *Carissa opaca* leaves on CCl_4 -induced damage in rat. *BMC Complement Altern Med* 11:48 www.biomedcentral.com/1472-6882/11/48
53. Zang Y, Guo J, Dong H, Zhao X, Zhou L, Li X, Liu J, Ni Y (2011) Hydroxysafflor yellow A protects against chronic carbon tetrachloride-induced liver fibrosis. *Eur J Pharmacol* 660(2-3):438–444
54. Wu S, Yue Y, Tian H, Li Z, Li X, He W, Ding H (2013) Carthamus red from *Carthamus tinctorius L.* exerts antioxidant and hepatoprotective effect against CCl_4 -induced liver damage in rats via the Nrf2 pathway. *J Ethnopharmacol*. <https://doi.org/10.1016/j.jep.2013.04.054>
55. Samoilik L, Lakic N, Mimica-Dukic N, Dakovic-Svajcer K, Bozin B (2010) Antioxidant and hepatoprotective potential of essential oils of (*Coriandrum sativum L.*) and caraway (*Carum carvi L.*) (*Apiaceae*). *J Agric Food Chem* 58:8848–8853
56. Shanmugasundaran R, Devi VK, Tresina PS, Maruthupandian A, Mohan VR (2010) Hepatoprotective activity of ethanol extracts of *Clitoria ternatea L.* and *Cassia angustifolia vahl* leaf against CCl_4 induced liver toxicity in rats. *IJRP* 1(1):201–205
57. Ilavarasan R, Mohideen S, Vijayalakshmi M, Manonmani G (2001) Hepatoprotective effect of *Cassia angustifolia Vahl*. *Indian J Pharm Sci* 63(6):504–507
58. Pradeep K, Mohan CVR, Anand KG, Karthikeyan S (2005) Effect of pretreatment of *Cassia fistula Linn.* leaf extract against subacute CCl_4 induced hepatotoxicity in rats. *Indian J Exp Biol* 43:526–530
59. Ahmed B, Khan S, Masood HM, Siddique AH (2008) Anti-hepatotoxic activity of Cichotybosome, a sequiterpene glycoside from seeds of *Cichorium intybus*. *J Asian Nat Prod Res* 10(3):218–223
60. Sadeghi H, Reza NM, Izadpanah G, Sohailla S (2008) Hepatoprotective effect of *Cichorium intybus* on CCl_4 -induced liver damage in rats. *Afr J Biochem Res* 2(6):141–144
61. Moselhy SS, Ali HHK (2009) Hepatoprotective effect of Cinnamon extracts against carbon tetrachloride induced oxidative stress and liver injury in rats. *Boil Res* 42:93–98
62. Bellassoued K, Hamed H, El Feki A, Ghrab F, Kallel R, van Pelt J, Lahyani A, Ayadi FM (2019) Protective effect of essential oil of *Cinnamomum verum* bark on hepatic and renal toxicity induced by carbon tetrachloride in rats. *Appl Physiol Nutr Metab* 44(6):606–618
63. Eidi A, Mortazavi P, Bazargan M, Zaringhalam J (2012) Hepatoprotective activity of *Cinnamom ethanolic* extract against CCl_4 -induced liver injury in rats. *EXCU J* 11:495–507
64. Karaca M, lihan F, Altan H, Him A, Tutuncu M, Ozbek H (2005) Evaluation of hepatoprotective activity of *Bergamot orange* in rats. *Eastern J Med* 10:1–4
65. Bhavsar SK, Joshi P, Shah MB, Santani DD (2007) Investigation of hepatoprotective property of *Citrus limon*. *Pharm Biol* 45(4):303–311
66. Molehin OR, Oloyede OL, Idowu KA, Adeyanju AA, Olowoyeye AO, Tubi Ol, Komolafe OE, Gold AS (2017) White butterfly (*Clerodendrum volubile*) leaf extract protect against carbon tetrachloride-induced hepatotoxicity in rats. *Biomed Pharmacother* 96:924–929
67. Pandey A, Bigoniya P, Raj V, Patel KK (2011) Pharmacological Screening of *Coriandrum sativum Linn.* for hepatoprotective activity. *J Pharm Bio allied Sci* 3(3):435–441
68. Sreelatha S, Padma PR, Umadevi M (2009) Protective effects of *Coriandrum sativum* extracts on carbon tetrachloride-induced hepatotoxicity in rats. *Food Chem Toxicol* 47:702–708
69. Li L, Zhou Y-F, Li Y-L, Wang L-L, Arai H, Xu Y (2017) *In vitro* and *in vivo* antioxidative and hepatoprotective activity of aqueous extract of *Cortex dictamni*. *World J Gastroenterol* 23(16):2912–2927
70. Lee G-H, Lee H-Y, Choi M-K, Chung M-K, Kim S-W (2017) Protective effect of *Curcuma longa L.* extract on CCl_4 -induced acute hepatic stress. *BMC Res Notes*:10:77. <https://doi.org/10.1186/s13104-017-2409-z>
71. Raja S, Nazeer Ahmed KFH, Kumar V, Mukherjee K, Bandyopadhyay A, Mukherjee PK (2007) Antioxidant effect of *Cytisus scoparius* against carbon tetrachloride treated liver injury in rats. *J Ethnopharmacol* 109:41–47
72. Balogun FO, Ashafa AOT (2016) Antioxidant and hepatoprotective activities of *Dicoma anomala Sond.* aqueous root extract against carbon tetrachloride-induced liver damage in Wistar rats. *J Tradit Clin Med* 36(4):504–513
73. Yeh Y-H, Hsieh Y-L, Lee Y-T (2013) Effects of yam peel extract against carbon tetrachloride-induced hepatotoxicity in rats. *J Agric Food Chem* 61:7387–7396
74. Beedimani RS, Shetkar S (2015) Hepatoprotective activity of *Eclipta alba* against carbon tetrachloride-induced hepatotoxicity in albino rats. *Inter J Basic Clin Pharm* 4(3):404–409
75. Sultana S, Ahmad S, Khan N, Jahangir T (2005) Effect of *Emblica officinalis (Gaertn)* on CCl_4 induced hepatic toxicity and DNA synthesis in Wistar rats. *Indian J Exp Biol* 43:430–436
76. Gupta G, More AS, Kumari RR, Lingaraju MC, Kumar D, Kumar D, Mishra SK, Tandan SK (2014) Protective effect of alcoholic extract of *Entada purpurea DC.* against CCl_4 induced hepatotoxicity in rats. *Indian J Exp Biol* 52:207–214
77. Pareek GA, Issarani R, Nagori BP (2013) Antioxidant hepatoprotective activity of *Fagonia schweinfurthii (Hadidi) Hadidi* extract in carbon tetrachloride induced hepatotoxicity in HepG2 cell line and rats. *J Ethnopharmacol* 150:973–981
78. Sharma M, Abid R, Ahmad Y, Nabi NG (2017) Protective effect of leaves of *Ficus carica* against carbon tetrachloride-induced hepatic damage in rats. *UK J Pharm Biosci* 5(1):6–11

79. Hsieh PC, Ho YL, Huang GJ, Huang MH, Chiang YC, Huang SS, Hou WC, Chang YS (2011) Hepatoprotective effect of the aqueous extract of *Flemingia macrophylla* on carbon tetrachloride-induced acute hepatotoxicity in rats through anti-oxidant activities. *Am J Chin Med* 39(2):349–365
80. Khattab HAH (2012) Effect of *Ginkgo biloba* leaves aqueous extract on carbon tetrachloride induced acute hepatotoxicity in rats. *Egypt J Hosp Med* 48:483–495
81. Nwidu LL, Oboma Yi, Elmorsy E, Carter WG (2018) Alleviation of carbon tetrachloride-induced hepatocellular damage and oxidative stress with a leaf extract of *Glyphaea brevis* (*Tiliaceae*). *J Basic Clin Physiol Pharmacol* 29(6):609–619
82. Duh P-D, Lin S-L, Wu S-C (2011) Hepatoprotection of *Graptotetalum paraguayense* E. Walther on CCl₄-induced liver damage and inflammation. *J Ethnopharmacol* 134:379–385
83. Zhang W, Zhang X, Xie J, Zhao S, Liu J, Liu H, Wang J, Wang Y (2017) Seabuckthorn berry polysaccharide protects against carbon tetrachloride-induced hepatotoxicity in mice via anti-oxidative and anti-inflammatory activities. *Food Funct.* <https://doi.org/10.1039/C7FO00399D>
84. Shahjahan M, Vani G, Shyamala Devi CS (2005) Protective effect of *Idigofolia oblongifolia* in CCl₄- induced hepatotoxicity. *J Med Food* 8(2):261–265
85. Khan RA, Khan MR, Ahmed M, Sahreen S, Shah AN, Shah MS, Bokhari J, Rashid U, Ahmed B, Jan S (2012) Hepatoprotection with a chloroform extract of *Launea procumbens* against CCl₄-induced injuries in rats. *BMS Compl Altern Med* 12:114 <http://www.biomedcentral.com/1472-6882/12/114>
86. Mohamed MA, Eldin IMT, Mohammed AH, Hassan HM (2016) Effects of *Lawsonia inermis* L. (*Henna*) leaves' methanolic extract on carbon tetrachloride-induced hepatotoxicity in rats. *J Intercult Ethnopharmacol* 5(1):22–26. <https://doi.org/10.5455/jice.20151123043218>
87. Hossain CM, Maji HS, Chakraborty P (2011) Hepatoprotective activity of *Lawsonia inermis* Linn, warm aqueous extract in carbon tetrachloride induced hepatic injury in Wistar rats. *Asian J Pharm Clin Res* 4(3):106–109
88. Sailar GU, Dudhereja AV, Seth AK, Maheshwari R, Shah N, Aundhia C (2010) Hepatoprotective effect of *Leucas cephalotes* speng on CCl₄ induced liver damage in rats. *Pharmacology Online* 1:30–38
89. Ulaganathan I, Divya D, Radha K, Vijayakumar TM, Dhanaraju MD (2010) Protective effect of *Luffa acurangula* (*Var*) amara against carbon tetrachloride-induced hepatotoxicity in experimental rats. *Res J Biol Sci* 5(9):615–624
90. Wills PJ, Asha VV (2006) Protective effect of *Lygodium flexuosum* (L.) Sw. extract against carbon tetrachloride-induced acute liver injury in rats. *J Ethnopharmacol* 108:320–326
91. Chaudhary A, Bhandari A, Pandurangan A (2011) Hepatic activity of methanolic extract of *Madhuca indica* on carbon tetrachloride- induced hepatotoxicity in rats. *Pharmacology Online* 1:873–880
92. Patel PK, Sahu J, Prajapati NK, Dubey BK, Alia A (2012) Hepatoprotective effect of ethanolic and hydro alcoholic leaf extract of *Madhuca indica* in carbon tetrachloride intoxicated rat. *Res J Pharmacol Pharmacodynamics* 4(5):311–314
93. Ramakrishna S, Geetha KM, Bhaskargopal PVVS, Ranjit Kumar P, Charan Madav P, Umachandar L (2011) Effect of *Mallotus philippensis* Muell-Arg leaves against hepatotoxicity of carbon tetrachloride in rats. *IJPSR* 2(2):74–83
94. Pramod K, Deval RG, Lakshmayya RSS (2008) Antioxidant and hepatoprotective activity of tubers of *Momordica tuberosa* Cogn. against CCl₄ induced liver injury in rats. *Indian J Exp Biol* 46:510–513
95. Bellassoued K, Hsouna AB, Athmouni K, Pelt J, Ayadi FM, Rebai T, Elfeki A (2018) Protective effects of *Mentha piperita* L. leaf essential oil against CCl₄ induced hepatic oxidative damage and renal failure in rats. *Lipids Health Dis* 17:9. <https://doi.org/10.1186/s12944-017-0645-9>
96. Patil K, Mall A (2012) Hepatoprotective activity of *Mentha arvensis* Linn. leaves against CCl₄ induced liver damage in rats. *Asian Pac J Trop Dis* 2(1):S223–S226
97. Rajendran R, Hemalatha S, Akasalai K, Madhakrishna CH, Vittal BS, Sundaram RM (2009) Hepatoprotective activity of *Mimosa pudica* leaves against carbon tetrachloride induced toxicity. *J Nat Prod* 2:116–122
98. Purkayastha A, Chakravarty P, Dewan B (2016) Evaluation of hepatoprotective activity of the ethanolic extract of leaves of *Mimosa pudica* Linn.in carbon tetrachloride induced hepatic injury in albino rats. *Inter J Basic Clin Pharmacol* 5(2):496–501
99. Jain A, Soni M, Deb L, Jain A, Rout SP, Gupta VB, Krishna KL (2008) Antioxidant and hepatoprotective activity of ethanolic and aqueous extracts of *Momordica dioica* Roxb. leaves. *J Ethnopharmacol* 115:61–66
100. Singh KG, Gupta GD (2012) Hepatoprotective and antioxidant activity of methanolic extract of flowers of *Nerium oleander* against CCl₄-induced liver injury in rats. *Asian Pac J Trop Med* 5(9):677–683
101. Abdus SS, Rahmat AK, Mushtaq A, Nawshad M (2016) Hepatoprotective role of *Nicotiana plumbaginifolia* Linn. against carbon tetrachloride-induced injuries. *Toxicol Ind Health* 32(2):292–298. <https://doi.org/10.1177/0748233713498448>
102. Ustuner D, Colak E, Dincer M, Tekin N, Donmez DB, Akyuz F, Colak E, Kolac UK, Entok E, Ustuner MC (2018) Post treatment effects of *Olea europaea* L. leaf extract on carbon tetrachloride-induced liver injury and oxidative stress in rats. *J Med Food* 20(0):1–6
103. Sikander M, Malok S, Parveen K, Ahmad M, Yadav D, Hafeez ZB, Bansal M (2013) Hepatoprotective effect of *Origanum vulgare* in Wistar rats against carbon tetrachloride-induced hepatotoxicity. *Protoplasma* 250:483–493
104. Brai BIC, Adisa RA, Odetola AA (2014) Hepatoprotective properties of aqueous leaf extract of *Persea Americana*, Mill (*Lauraceae*) 'avocado' against CCl₄-induced damage in rats. *Afr J Tradit Complement Altern Med* 11(2):237–244
105. Ezat M, Okba MM, Ahmed SH, El-Banna AP, Mohamed SO, Ezzat SM (2020) In-dept hepatoprotective mechanistic study of *Phyllanthus niruri*: *in vitro* and *in vivo* studies and its chemical characterization. *PLoS One* 15(1):e0226185. <https://doi.org/10.1371/journal.pone.0226185>
106. Khalaf-Allah AM, El-Gengahi SE, Hamed MA, Zahran HG, Mohammed MA (2016) Chemical composition of golden berry leaves against hepato-renal fibrosis. *J Diet Suppl* 13(4):378–392
107. Abdel Raouf GF, Said AA, Mohamed KY, Gomaa HA (2020) Phytoconstituents and bioactivities of the bark of *Pleiogynium timorensis* (DC) Leenh (*Anacardiaceae*). *J Herbmed Pharmacol* 9(1):20–27
108. Jayakumar T, Ramesh E, Geraldine P (2006) Antioxidant activity of the oyster mushroom, *Pleurotus ostreatus*, on CCl₄-induced liver injury in rats. *Food Chem Toxicol* 44:1989–1996
109. Mahmud ZA, Bachor SC, Qais N (2012) Antioxidant and hepatoprotective activities of ethanolic extracts of leaves of *Preruna esculenta* Roxb against carbon tetrachloride-induced liver damage in rats. *J Young Pharm* 4(4). <https://doi.org/10.4103/0975-1483-104360>
110. Syed SN, Rizvi W, Kumar A, Khan AA, Moin S, Ahsan A (2014) *In vitro* antioxidant and *in vivo* hepatoprotective activity of leave extract of *Raphanus sativus* in rats using CCl₄ model. *Afr J Tradit Complement Altern Med* 11(3):102–106
111. Kalegari M, Gemin CAB, Araujo-silva N, de Brito NJ, Lopez JA, Tozetto S, Almeida M, Migue IMD, Stien D, Miguel OG (2014) Chemical composition, antioxidant activity and hepatoprotective potential of *Rourea induta* planch (*Connaraceae*) against CCl₄-induced liver injury in female rats. *Nutr* 30:713–718
112. Rao GMM, Rao CV, Pushpangadan P, Shirwaikar A (2006) Hepatoprotective effects of rubiadin, a major constituent of *Rubia cordifolia* Linn. *J Ethnopharmacol* 103:484–490
113. Tukaappo NK, Londonkar RL, Nayaka HB, Kumar SCB (2015) Cytotoxicity and hepatoprotective attributes of methanolic extract of *Rumex vesicarius* L. *Biol Res* 48(19). <https://doi.org/10.1186/s40659-015-0009-B>
114. Sun ZL, Gao GL, Xia YF, Qiao ZY (2011) A new hepatoprotective saponin from *Semen celosia cristatae*. *Fitoterapia* 82(4):591–594
115. Shahjahan M, Sabitha KE, Jainu M, Shyamala Devi CS (2004) Effect of *Solanum trilobatum* against carbon tetrachloride induced hepatic damage in albino rats. *Indian J Med Res* 120:194–198
116. Gupta RK, Hussain T, Panigrahi G, Das A, Singh GN, Sweety K, Faiyazuddin MD, Rao CV (2011) Hepatoprotective effect of *Solanum xanthocarpum* fruit extract against CCl₄-induced acute liver toxicity in experiments. *Asian Pac J Trop Med* 4(12):964–968
117. Nwidu LL, Elmorsy E, Oboma Yi, Carter WG (2018) Hepatoprotective and antioxidant activities of *Spondias mombin* leaf and stem extracts against carbon tetrachloride-induced hepatotoxicity. *J Taibah Univ Med Sci* 13(3):262–271
118. Kokhdan EP, Ahmadi K, Sadeghi H, Dadgery F, Danaei N, Aghamaali MR (2017) Hepatoprotective effect of *Stachys pilifera* ethanol extract

- in carbon tetrachloride-induced hepatotoxicity in rats. *Pharm Biol* 55(1):1389–1393
119. Deng J-S, Chang Y-S, Wen C-L, Liao J-C, Hou W-C, Amagaya S, Huang S-S, Huang G-J (2012) Hepatoprotective effect of the ethanol extract of *Vitis thunbergii* on carbon tetrachloride-induced acute hepatotoxicity in rats through anti-oxidative activities. *J Ethnopharmacol* 142:795–803
120. Song A, Ko HJ, Lai MN, Ng LT (2011) Protective effects of Wu-Liang-Shen (*Xylaria nigritipes*) on carbon tetrachloride-induced hepatotoxicity in mice. *Immunopharmacol Immunotoxicol* 33(3):453–460
121. Oke GO, Abiodun AA, Imafidon CE (2019) *Zingiber officinale* (Roscoe) mitigates CCl_4 -induced liver histology and biochemical derangements through antioxidant, membrane-stabilizing and tissue-regenerating potential. *Toxicol Rep* 6:416–425
122. Shen X, Tang Y, Yang R, Yu L, Fang T, Duan J (2009) The protective effect of *Zizyphus jujube* fruit on carbon tetrachloride-induced hepatic injury in mice by anti-oxidative activities. *J Ethnopharmacol* 122:555–560
123. Chang L, Xu D, Zhu J, Ge G, Kong X, Zhou Y (2020) Herbal therapy for the treatment of acetaminophen-associated liver injury: recent advances and future perspectives. *Front Pharmacol* 11:313. <https://doi.org/10.3389/fphar.2020.00313>
124. Stickel F, Shouval D (2015) Hepatotoxicity of herbal and dietary supplements: an update. *Arch Toxicol* 89(6):851–865
125. Janghel V, Patel P, Chandel SS (2019) Plants used for the treatment of icterus (jaundice) in Central India: A review. *Ann Hepatol* 18:658–672
126. Zhu J, Chen M, Borlak J (2019) The landscape of hepatobiliary adverse reactions across 53 herbal and dietary supplements reveals immune-mediated injury as a common cause of hepatitis. *Arch Toxicol* 94(1):273–279
127. Shakya AK (2020) Drug-induced hepatotoxicity and hepatoprotective medicinal plants: a review. *Indian J Pharm Edu Res* 54(2):234–247

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Submit your manuscript to a SpringerOpen® journal and benefit from:

- Convenient online submission
- Rigorous peer review
- Open access: articles freely available online
- High visibility within the field
- Retaining the copyright to your article

Submit your next manuscript at ► springeropen.com